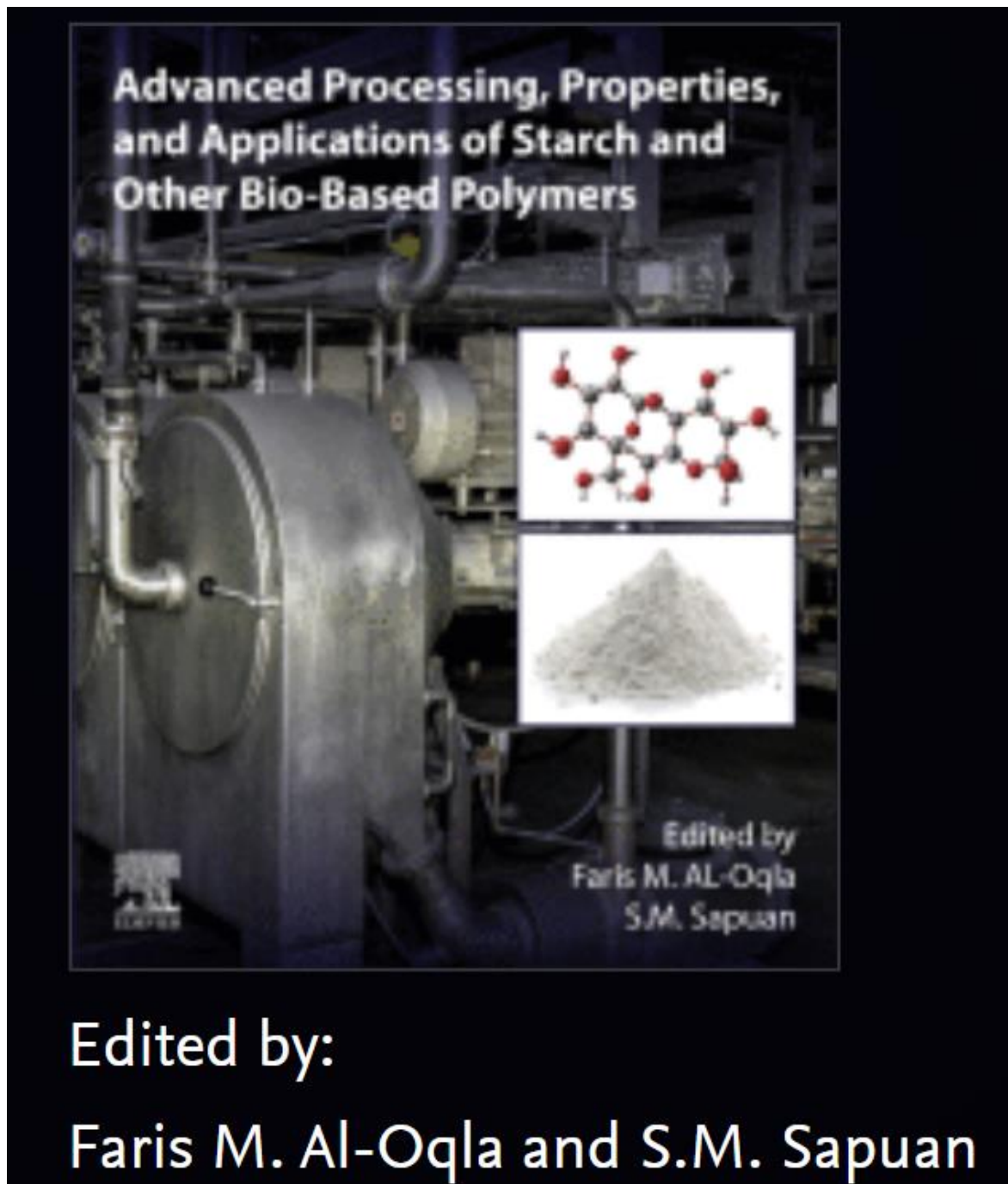



# Properties and Characterization of PLA, PHA, and Other Types of Biopolymer Composites



CHAPTER Book



# Advanced Processing, Properties, and Applications of Starch and Other Bio-Based Polymers

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50 Hampshire Street, 5th Floor, Cambridge, MA 02139, United States

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#### Library of Congress Cataloging-in-Publication Data

A catalog record for this book is available from the Library of Congress

#### British Library Cataloguing-in-Publication Data

A catalogue record for this book is available from the British Library

ISBN: 978-0-12-819661-8

For information on all Elsevier publications visit our website at  
<https://www.elsevier.com/books-and-journals>

Publisher: Matthew Deans  
Acquisitions Editor: Edward Payne  
Editorial Project Manager: Charlotte Rowley  
Production Project Manager: Kiruthika Govindaraju  
Cover Designer: Alan Studholme

Typeset by TNQ Technologies



Full paper silahkan di

[https://drive.google.com/file/d/1gHDJtnfzZXd8JL7Ju8Ljv5F0B\\_rtONXt/view?usp=sharing](https://drive.google.com/file/d/1gHDJtnfzZXd8JL7Ju8Ljv5F0B_rtONXt/view?usp=sharing)

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# Properties and Characterization of PLA, PHA, and Other Types of Biopolymer Composites

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## 1 INTRODUCTION

The production of petroleum-based plastic had increased tremendously, reaching about 350 million tons annually (Garside, 2019). According to Rochman et al. (2013), the earth will accumulate with about 33 billion tons of plastic wastes by 2050, if current rates of consumption continue. Besides that, inappropriate usage and disposal of plastics waste would lead to substantial pollution of both terrestrial and marine ecosystems. The world produces about 350 million tons of plastic waste annually, and surprisingly, only 9% of this waste has been recycled (UNEP, 2018). Moreover, it had been estimated that each year at least 8 million tons of plastic waste go to the rivers and oceans, and some of them often decompose into small microplastics that end up stopping in our food chain (UNEP, 2018). Because of the environmental challenges aiming to reduce this environmental impact, many researchers have formulate eco-friendly and biodegradable composites polymer to replace conventional petroleum-based polymer.

Recently, many countries have banned petroleum-based plastics because of the huge volume of plastic waste that harmfully affects the ecosystem, wildlife, and environment (Aisyah et al., 2019; Asyraf et al., 2020; Atikah et al., 2019; Norizan et al., 2020; Nurazzi et al., 2019a). These nondegradable plastic are responsible for the “white pollution” worldwide. The “white pollution,” including plastic bags, plastic bottles, plastic silverware, and other materials that are made from the plastic, kills the wildlife, marine life, and avifauna and degrades the quality and features of the environment on the Mother Nature (Thiagamani et al., 2019). Remarkably, Malaysia is the foremost country

in Southeast Asia region to take courageous act to confront “white pollution.” The Government of Malaysia has broadcasted that the government will ban single-use plastic by year of 2030 (UNEP, 2018). Although Malaysia is a bit behind when it comes to enacting against single-use plastics, nevertheless according to New Strait Times, Federal Territories of Malaysia has announced that from March 2019, a pollution charge of 20 cent imposed for a single plastic bag. Therefore, customers will either have to pay 20 cent for a reusable bag or bring their own bags. Besides Malaysia, others countries such as Kenya, China, Rwanda, Uganda, Ireland, South Africa, Morocco, Taiwan, India, France, and Canada have already forbid and eliminated completely the use of single-use plastic and plastic bags.

Therefore, in order to cater this problems, biodegradable polymers were introduced. Biodegradable polymers are one of the potential solutions to the problems associated with discarded wastes (Abral et al., 2020a,b; 2019a; Atiqah et al., 2019; Ilyas et al., 2018, 2017; Nurazzi et al., 2019b). This is due to their fast degradation by the action of naturally occurring microorganisms in the environment (Ilyas et al., 2018a). Biodegradable polymer can be produced via (1) bio-based (i.e., polyhydroxyalkanoates [PHA], starch, protein, polylactic acid [PLA], chitin, chitosan, polybutylene succinate [PBS], and cellulose) and (2) fossil- or petroleum-based (i.e., polycaprolactone [PCL], poly(vinyl alcohol), and poly(butylene adipate-co-terephthalate) materials.

Generally, polymer is made up of long chain of polymer with many repeated subunits. Both man-made

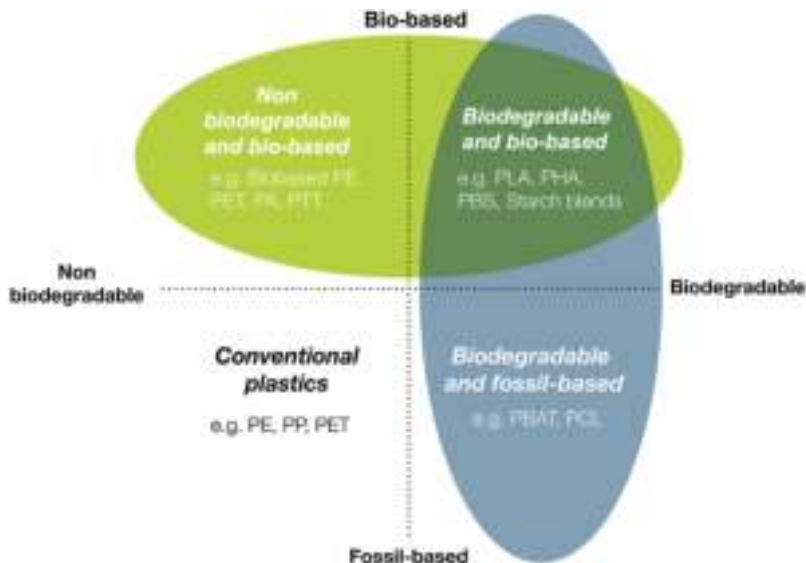
(synthetic) and natural polymers play important and ubiquitous roles in daily life. This is because of their various unique properties (Ilyas et al., 2018a). They are made by the process of condensation and addition polymerization reactions. They can be categorized either as thermosetting or thermoplastic polymers. “Bio-based plastic” can be defined as a plastic that are made from natural resources and it is renewable, whereas “biodegradable plastic” is referred on how the plastics behave at end of its life. The illustrations of bioplastic is displayed in Fig. 8.1.

Biodegradable plastic is a material that is decomposed naturally when introduced in the environment by the action of living organisms, usually microorganisms. It is eco-friendly compared with conventional plastics. Besides that, this plastic is commonly produced via microorganism, renewable raw materials, petrochemicals, or combination of all three. There are a lot of materials that can be used to make biodegradable and bio-based plastics such as plants, starches (cassava, corn, potato, sugar palm, yam bean, pea, wheat tapioca, bengkuang), peels from citrus fruits, and corn oil (Atikah et al., 2019; Halimatul et al., 2019a,b; Jumaidin et al., 2019a–c; Syafri et al., 2019). Biodegradable and bio-based plastic offer a material that is made from natural resources, therefore the risk of breaking down these material are much fewer compared with conventional petroleum-based plastic. Fig. 8.2 shows some of the list of the advantages and disadvantages of utilizing biodegradable plastic.

Biodegradable plastic is often reflected as a savior product. Shifting to these materials would decrease the levels of greenhouse gas emission, reduce carbon dioxide levels, reduce energy of manufacturing, reduce amount of waste produced, and create new plastic industries. However, there is also the potential for biodegradable plastic to create more pollution. This is because biodegradable plastic is difficult to be decomposed in ocean water like it would during composting in soil. This plastic will either float on the ocean or river surface like other conventional plastic or more worse it create microplastic that is harmful to marine life, sea life, or ocean life. Therefore, shifting to this biodegradable plastic is not a final solution that will resolve our pollution problems. More research must be done to develop total biodegradable plastic that can be degraded fully in every condition. The advantages and disadvantages of biodegradable plastic provide opportunity to reduce human dependence in fossil fuels especially in reliance on crude oil or petroleum. There are several types of biodegradable and bio-based polymer such as PHA, PLA, starch, protein, chitin, chitosan, and PBS (Azammi et al., 2020). Therefore, this chapter focuses on providing an overview of recent advancements on biodegradable bio-based polymer for various application.

## 2 POLYHYDROXYALKANOATES

PHA are a kind of degradable plastics. It has good environmental effects compared with petroleum-based



**FIG. 8.1** Conventional petroleum-based plastics and bioplastics are that made up of biodegradable polymers (Ilyas and Sapuan, 2020).



### Advantages

- Can be broken down by naturally occurring microorganism
- Reduces greenhouse gas emission levels
- Reduces carbon dioxide levels
- Does not release other harmful products after disposal
- Uses low energy during the production cycle
- Reduces the amount of waste we produce
- Current use of petroleum can be channeled to other needs
- Can mix and blend with conventional plastic
- Could create new export industries
- Could establish a new business opportunity and marketing platform
- Can degrade faster under certain conditions

### Disadvantages

- Disposal of bioplastic waste must be through certain procedures
- Requires the some certain environmental conditions such as UV light, temperature and humidity) to associated with their disposal
- The costs of pesticides and herbicides are not taken into account during the manufacturing of biodegradable plastic
- The utilization of biodegradable plastics would reduce the number of plastic that can be recycling due to their limited properties
- High capital cost
- Requires croplands to produce items
- Some of the bioplastics only reduce their size to microsize plastic during decomposition process, but this does not solve the current ocean pollution problems, besides contaminated microplastics could expose marine organism to high concentrations of toxins
- May produce methane in landfills

**FIG. 8.2** Advantages and disadvantages of biodegradable plastics.

polymer. PHA is a natural biopolymer material developed rapidly in recent 20 years. PHA are polyesters that were synthesized in nature by many microorganisms, including bacterial fermentation of lipids and sugars.

PHA has good mechanical, thermal processing, biodegradability, and biocompatibility properties. It can be used in agricultural materials, biomedical materials, and packaging applications. Recently, this polymer has attract huge attention in field of biomaterials.

Besides that, the blending polymer of starch and aliphatic polyester from sustainable and renewable resources to produce biodegradable plastics such as garbage bags and other products has been successfully studied and applied in American and European countries. One of the company named Novamont in Novara, Italy, had promoted a bioeconomy model based on the efficient use of resources and on territorial regeneration (Sahay and Ierapetritou, 2009). This company sets up biorefineries for the production of bioplastics and bio-products of renewable resources conceived to protect environment, ecology, and wildlife. In 2018, Novamont had announced the opening of a revamped

Mater-Biopolymer plant south of Rome to produce MATER-BI. This plant significantly boosting production capacity from 120,000 tons per year to 150,000. Remarkably, MATER-BI is widely used in Europe and the United States.

Polyhydroxyalkyl fatty acid esters (PHA) are biosynthetic polyesters consisting of a series of different repetitive unit structures. PHA has many interesting properties and has been utilized in many applications such as medical applications, fishing lines, and plastics bag. The biosynthesis of PHA monomer depends on many aspects, including the types of carbon sources on which biological growth depends, the types of metabolic pathways through which organisms convert these carbon sources into PHA monomers, and the substrate specificity of enzymes involved in PHA synthesis (Pérez-Arauz et al., 2019).

## 2.1 Properties of Polyhydroxyalkanoates

PHA is a kind of thermoplastic material with high degree of result. Its physical properties and chemical structure are basically similar to polypropylene (PP) and

polyethylene (PE), and it is capable of drawing, pressing film, injection molding, etc. PHA is a kind of intracellular carbon source and energy storage, which can be naturally decomposed and utilized by many microorganisms and is biodegradable.

Soil burial experiments showed that PHA films with thickness of 0.07 mm could be degraded basically in about 6 weeks. The intracellular degradation of PHA was mainly through the formation of monomers or dimers by PHA depolymerase. There were two mechanisms of extracellular degradation, one was the automatic hydrolysis of melting bonds without supervision, and the other was decomposed by depolymerization of extracellular PHA. For example, as shown by Mergaert et al., 295 microorganisms have been found to degrade PHA in soil. In addition to the main characteristics of biodegradability, PHA has special properties such as bio-PHAe transport, optical activity, piezoelectricity, foam resistance, low permeability, etc. It can be widely used in industry, agriculture, medicine, scientific research, and other fields (Lu et al., 2009). In the PHA family, more than 90 different monomers have been discovered, except for a small number of 4-hydroxybutyric acid, 4-hydroxyvaleric acid, and 5-hydroxyvaleric acid, most of which are 3HA. At most, the fermentation mechanism and its properties are well known.

## 2.2 Advantages of Polyhydroxyalkanoates

PHA (and bioplastics in common) are exceptionally appealing materials for three essential reasons: they can be made from renewable sources, they can biodegrade, and they are biocompatible. To the primary point, it is exceptionally energizing that researchers are finding ways to gather and utilize fabric from sources like microbes amalgamation (PHA) and corn or sugarcane (other bioplastics like PLA). Already, crops had to be redirected for the production of bioplastics, but within the final decade or so, there has been a center on utilizing squander materials (such as banana peels, potato peelings, etc.) to deliver bioplastics instep. By utilizing squander items, utilization of rare assets can be maximized.

Biodegradability is the other key viewpoint that creates PHA an awfully promising material. Environmental contamination could be a hot subject with vital suggestions; ordinary petrochemical-based plastics have been at the exceptionally center of the controversy, fundamentally since they are so broad and do not degrade effectively. For occasion, pictures of marine creatures choked by or ingesting different plastics show the stark reality of contamination. Of course, fossil fuel-based plastics have

done much to move forward the material lives of people, but when it comes to contamination, modern developments in biodegradability are exceptionally welcome.

Since PHA are biocompatible (which implies not destructive to living tissue), they can and have been used in an assortment of therapeutic and surgical applications. Looking forward, the potential moreover exists that PHA will be included in “wearable” inner innovation applications.

PHA can reduce the landfill needed to bury the plastic waste as the period time needed for PHA to biodegrade is fast and reliable, which reduces the impact to environment. Besides, biopolymers can decrease the natural affect derived from plastic waste transfer due to the truth that the biodegradation time for biopolymers within the land surface beneath standard conditions is roughly 2 months (Hassan et al., 2013).

PHA have the advantages to biodegrade in most of the situation, not only on land surface but also in water area due to the special process undergone during the biodegradation process. PHA can be biodegraded under both anaerobic and aerobic conditions by PHA degraders as shown in most situation, including the marine environment (Shah et al., 2008).

## 2.3 Application of Polyhydroxyalkanoates

Bacterial plastics have gained a very high attention worldwide and globally as the better choice in packaging materials. The applications include in medical devices, skin care, personal hygiene products, plastic packaging, and utilized in agricultural field as mulching films. It is an alternative option for conventional plastics that are being used in agriculture field, which are nonbiodegradable and nonenvironment friendly. This bacterial plastics are the better substitution for those examples mentioned previously because these conventional plastic are single-use application and the demand amounts are enormous, thus creating a lot of plastic waste. According to Chen (2010), the application of PHA is not only limited to bioplastics application, however, PHA have also been make full use as implant biomaterials, biofuels, fine chemicals, medicines, and for regulating bacterial metabolism as well as enhancing the quality of industrial microorganisms.

The scope of application was made possible by further chemically modifying the PHA's functional groups. The utilization of PHA in various industries has grown considerably wide. Currently, aluminum is used as cover for cupboard to prevent water from entering the product. Hence, to overcome this problem, PHA latex can be utilized to cover cardboard to make

the surface of product water-resistant. Besides that, only small amount of PHA is required for this purpose, and thus it works out to be cost-effective alternative (Patricia et al., 2007).

Nonetheless, PHA is also extensively use in the tissue engineering. Tissue engineering involves the use of a tissue scaffold for the formation of new viable tissue for a medical purpose. Therefore, suitable material for this application must have properties such as support cell growth, allow tissue ingrowth, guide and organize the cells, degrade to nontoxic products, and biocompatibility. Thus, according to these criteria, PHA is seem completely fit into these criteria and make it the best candidate as the suitable material to be used in tissue engineering application.

Furthermore, PHA also can be utilized in controlled drug release systems. The capability of this material to work with a suitable host response and biodegradability properties make it useful for drug delivery in the medical field. Besides that, many researches have been conducted and stated that various variety of monomers can be added into PHA for modification. These modification would result in numerous changes of physical properties that range from strong elastomers to highly crystalline materials. The rate of decomposition can be indirectly controlled by precisely controlling the monomer composition of PHA. Catalyst reaction of the enzymatic degradation is normally done by bacterial PHA depolymerase.

Metabolix, a US-based company, blend P(3HB) and poly(3-hydroxyoctanoate) to produce a new compound of PHA and marketed their product in the market. This newly formed PHA compound is an elastomer that has been ratified by the Food and Drug Administration for usage as food additives. This example shows that the range of PHA application has been expanded widely as they could even found in the food industries.

Following by the application in food additive, the application of PHA can be found in the electronic industries as well. This function is possible due to the PHA piezoelectric nature. The electronic components or parts that can be produced by PHA are stretch and acceleration measuring instruments, shock wave sensors, gas lighters, lighters, material testing, pressure sensors for keyboards, oscillators: for atomization of liquids and ultrasonic therapy, loudspeakers, headphones, and acoustics: sound pressure measuring instruments, ultrasonic detectors, and microphone.

Last but not least, polystyrene (PS) waste was renowned as the waste materials that takes the longest degradation time or might be nondegradable. However,

with the advanced scientific exploration, researchers were able to devise a novel way to alter the abundant PS waste into PHA biopolymer using combination of pyrolysis. This process is cost-effective and efficient and can be one of the ways to optimize the use of PS residues.

### 3 POLYLACTIC ACID

World issues such as environmental, economic, and health issues have been constantly pushing scientists, researchers, and manufacturers to slowly replace the extensive use of plastics in the current world. Most plastics produced today are made from nonrenewable resources such as petroleum. Unlike any other plastics, PLA is made up from renewable resources such as corn starch or sugar cane (Siakeng et al., 2019). PLA is known as bioplastic because it is obtained from biomass. Not only that PLA is biodegradable, it also has attributes like PS, PE, or PP. Due to its similar production process, it can be produced easily from the existing production plant for petrochemical industry plastics. This advantage makes it cost effective to be utilized. PLA is mainly produced through condensation and polymerization processes. Ring-opening polymerization process is a procedure that uses metal catalyst mix with lactide to form a bigger PLA molecules. Similarly, the condensation process goes through the same procedure with different temperature and produce different by-products (Singhvi et al., 2019). There are few types of PLA, e.g., racemic PLLA, regular PPLA, PDLA, and PDLA. All of them are having different characteristics yet similar because they are derived from renewable sources. PLA is a thermoplastic polyester which means that it can be heated to their melting point at around 150–160°C, cooled, and heated again without any degradation. In contrast, the thermoset plastic can be heated only once and it is irreversible. Due to its characteristics, PLA has been the most studied and utilized biodegradable plastic in the human history. It is replacing conventional petrochemical-based polymers slowly and becomes the leading biomaterial for medical application as well as in other plastic industries (Farah et al., 2016).

#### 3.1 Advantages of Polylactic Acid

PLA is tremendously used in research and daily life as biocompatible polymer due to properties without toxic or carcinogenic effects for human body (Rasal et al., 2010). For quite a long time, we have been cautioned of the hazardous synthetic substances that escaped when common plastics are burned. In biological

perspective, PLA plastics do not develop these poisonous gas if ended up in the event of burning instead of finding their path to a composting facility operate in large scale.

Most of the polymers that exist nowadays are derived from nonrenewable resources especially petrochemical, which are just accessible in limited quantity all through the world. In the end, these fossil resources will be used up. Production of synthetic polymers, as well as to get rid of it by burning, will create a lot of CO<sub>2</sub>, which contributes to the global warming (Pang et al., 2010).

PLA, which is produced from corn, is a resource that can be restored every year (Singhvi et al., 2019). PLA are getting more attention commercially since they are produced from corn-based starch, sequester critical amounts of CO<sub>2</sub> in respect to petrochemical-based materials, save energy, and degrade in a short time. Researchers have shown that PLA demonstrates lower fossil resources utilization, which reduces the risk of summer smog and global warming.

PLA will bring a lot of benefits such as its preparation from lactide monomer which can get from a renewable source in agriculture field (Gewin, 2003; Sawyer, 2003), low quantities of CO<sub>2</sub> used up (Dorgan et al., 2001), the contribution in reducing amount of landfill and developing economies of farm, and lastly better mechanical properties compared to PS and PET (Auras et al., 2005).

### 3.2 Disadvantages of Poly(lactic Acid)

Table 8.1 give a summary on the degradation rate of some synthetic polyesters. It show that PLA is slower than PGA, PLGA, and PCLA copolymers in term of degradation rate. The reason behind this condition is

**TABLE 8.1**  
Biodegradation Rate of Some Polyesters (Ikada and Tsuji, 2000; Zhu et al., 1991).

Polymers	Molecular Mass (kDa)	Degradation Rate
PLLA	100–300	(SC) 50% in 1–2 years
PGA	–	(C) 100% in 2–3 months
PLGA	40–100	(A) 100% in 50–100 years
PCL	40–80	(SC) 50% in 4 years
PCLA	100–500	(A) 100% in 3–12 months
PTMC	14	(SC) 9% in 30 weeks

the poor properties that make the water diffusion difficult to the semicrystallinity of PLA.

To some certain extent, despite the fact that PLA is biodegradable, the duration taken will be so long. As indicated by Elizabeth (2006), PLA may well separate into its constituent parts, carbon dioxide and water in a controlled composting condition, that is, increase the surrounding temperature to 140°F. In any case, it will take much more time in a fertilizer container or in a landfill stuffed so firmly that no light and little oxygen are accessible to aid the procedure.

It is regretting to say that, most PLA plastic will not break down into natural components in any composting pile appearing in courtyard. Rather, these items should be transported to a commercial composting facility. In any case, as the business develops, we trust that the facility for commercial composting will tag along. Discarding PLA plastic items in a landfill would be considered as a suicide alternative.

### 3.3 Application of Poly(lactic Acid)

There are three main applications for PLA plastics, which are domestic, medical, and packaging and 3D printing applications. Table 8.2 shows the applications of PLA and its usages.

## 4 STARCH

Starch-based biopolymer films have been extensively utilized in medicine as well as food packaging application, in which the biofilm should be edible in many cases, such as applications in medicine capsules, candy wrappers, etc. This type of biofilm has a potential to be used for controlling water permeability, as a barrier for volatile compounds and gases, and to maintain the food freshness. However, starch-based biopolymer has poor mechanical properties and high water vapor permeability. To overcome this drawback, starch-based biopolymers are (1) blend with other polymers, (2) reinforced with plasticizer, particle, or fiber fillers, or (3) modified to starch structure (Ilyas et al., 2020a,b, 2019a–d, 2018a–d; Sanyang et al., 2018). Nevertheless, the reinforcement of filler or plasticizer has some problems related to safety issues. Therefore, the selection of filler, plasticizer, polymer blend, and chemical used to modified starch must be nonallergic, nontoxic, fully biodegradable, and digestible and can be consumed by living things.

### 4.1 Properties of Starch

Physically, most native starches are semicrystalline, having a crystallinity of about 20%–45% (Abral et al., 2019b). The short-branched chains in the amylopectin

**TABLE 8.2**  
The Application of PLA Plastics (Jamshidian et al., 2011).

Number	Applications	Usages
1	Domestics	Plates and saucers, cups, cutlery, fruit juices, fresh water, sports drinks, cold drink cups, transparent food containers, foodware, dairy containers, jelly and jam container, and edible oils container.
2	Medical	Medical devices such as plates, rods, pins, and screws.
3	Packaging	Vegetable bags, candy twist wrap, lidding film, salad, blister packaging, window envelope film, label film, shrink wrap material, and other packaging applications.
4	FDM machines (3D printing)	3D printable filament, lost PLA casting for molten metal, and other 3D printing medical device prototypes (both biodegradable and permanent).

are the main crystalline component in starch granular. Crystalline regions exist in the form of double helices with a length of  $\sim 5$  nm. Besides that, the amylopectin segments in the crystalline regions are all parallel to the axis of the large helix. The molecular weight of amylose is about 100 times lower than that of amylopectin. Moreover, the ratio of amylose to amylopectin much depends on the age and source of the starch. In addition, the ratio of amylose and amylopectin can also be controlled by the extraction process method used. Starch granules also contain small amounts of lipids and proteins. Fig. 8.3 shows the chemical structures and physical diagram illustration of amylopectin starch and amylose starch (Generalic, 2019). Moreover, thermoplastic starch (TPS) is formed by disrupting the ordered structures within the starch molecular. Heating process is required along with shear force to disrupt the starch granules. This shear forced and heating process would cause swelling and nonirreversible transition of amorphous regions in the presence of plasticizer, under certain condition

(Sanyang et al., 2016). Table 8.3 shows the chemical composition of commercial starches. Besides that, from Table 8.4, it can be observed that the highest amylose content is sugar palm starch (SPS). Table 8.5 shows the SPS properties in comparison with sago starch. The mechanical properties of SPS biofilm was observed higher compared with sago starch biofilm. Therefore, SPS biofilms have higher potential to be utilized as bio-based packaging.

## 4.2 Advantages and Disadvantages of Starch Biopolymer

The development of novel starch-based polymer materials using renewable resources became hot topic among the researchers to overcome the environmental problems caused by plastic wastes. Starch can be extracted from sugar palm tree, tapioca, wheat, tapioca, potato, cassava, bengkuang, and maize (Jumaidin et al., 2019a,b, 2020). Generally, such material is stored in plants tissues as one-way carbohydrates. It is made up of glucose and can be attained by melting starch. However, this polymer is not available in animal tissues. Starch possess several advantages such as abundance, renewability, easy availability, biodegradability, ease process, and cheap. Native starch-based biopolymer possess many disadvantages, such as high thermal degradation rates, low mechanical properties, high water-barrier properties and processability (Hazrol et al., 2020; Nazrin et al., 2020). The disadvantages of starch-based biopolymer are listed in Fig. 8.4. Current technique that is being used to process starch nowadays is solution technique. Solution casting technique is the easiest and most widely utilized at laboratory scale compared with other method such as hot press. However, this technique cannot be conducted at large-industrial scale as it encompasses too long drying time processed. Therefore, in order to overcome this problem, the fabrication of starch-based polymer by thermoplastic treatments (reactive extrusion, foaming extrusion, film/sheet extrusion, and injection molding) could be considered.

## 4.3 Application of Starch Biopolymer

Bio-based and biodegradable starch-based biopolymers have an extensive range of applications such as mulching film horticultural crops, drop ceiling tiles, pharmaceutical, biomedical, corrugated board adhesives, paper, horticulture, agriculture, consumer electronics, automotive, textiles, and packaging (Spaccini et al., 2016; Tan et al., 2016). Table 8.6 summarized the starch-based biopolymer, its manufacturing technique, and applications.

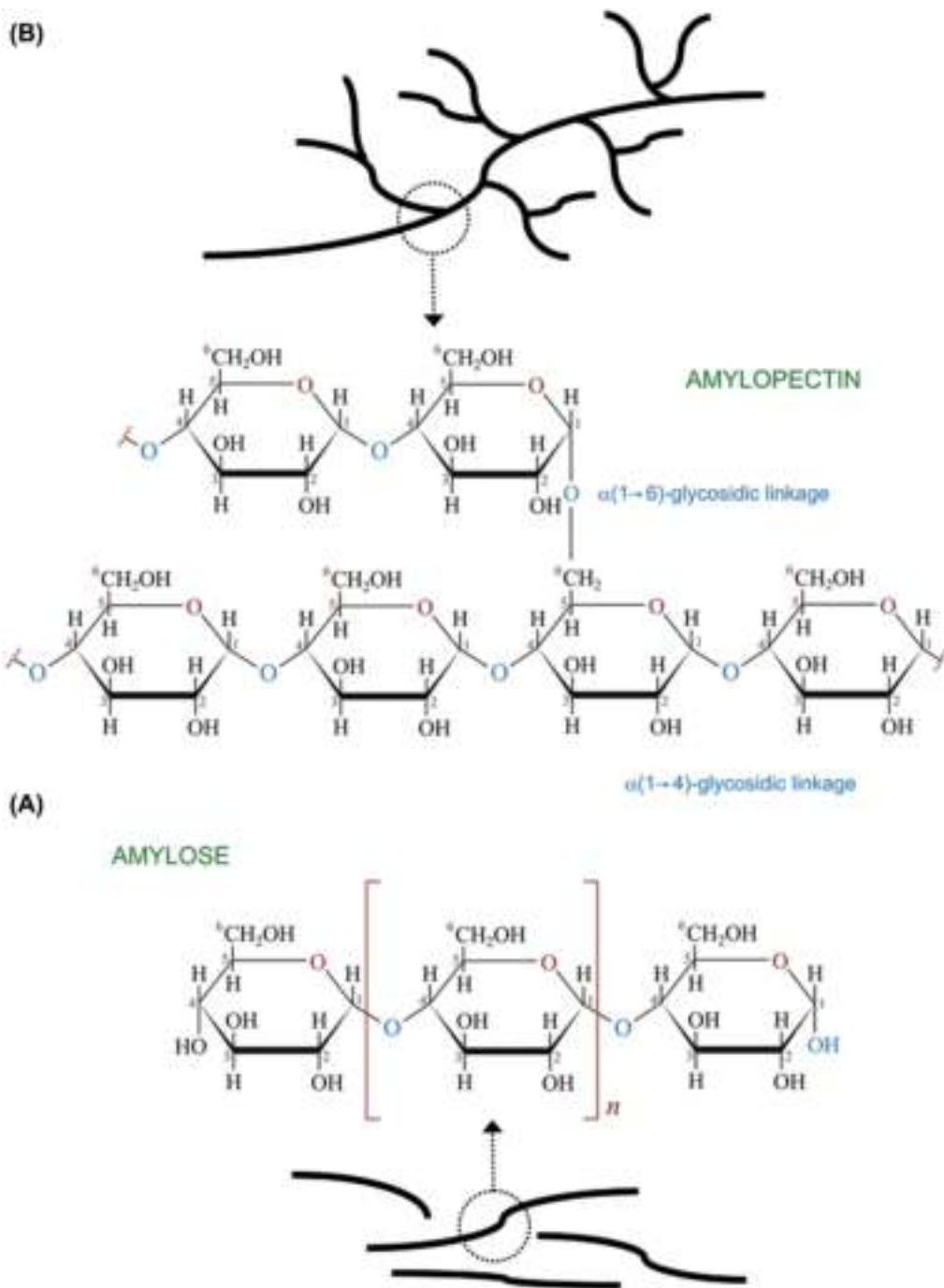


FIG. 8.3 Chemical structures and physical schematic representation of (A) amylose starch and (B) amylopectin starch.

**TABLE 8.3**  
The Chemical Composition of Commercial Starches (Sanyang et al., 2016).

Starch	Density	Ash (%)	Amylose (%)	Water Content (%)
Wheat	1.44	0.2	26–27	13
Tapioca	1.446 –1.461	0.2	17	13
Maize	1.5	0.1	26–28	12–13
Potato	1.54 –1.55	0.4	20–25	18–19
Sago	–	0.2	24–27	10–20
Sugar palm starch	1.54	0.2	37.60	15

## 5 PROTEIN

### 5.1 Properties of Protein Biopolymer

Protein is considered as one of the most plentiful biological macromolecules in cells, occurring in an extensive variety of species and ranging in size from relatively small peptides to polymers with high molar mass. These molecules exhibit diverse biological functions (Durán et al., 2011), providing structure or biological activity in animals or plants. Besides that, proteins are well-known and compared with other macromolecules due to their structure that is based on approximately 20 amino acid monomers, rather than just a few or even one monomer, such as glucose in the case of cellulose and starch. Most proteins contain 100–500 amino acid residues (Fennema, 1985; Haghpanah et al., 2009).

The functional variety of proteins basically arises from their chemical structure. Depending on the sequential order of the amino acids, the protein will

**TABLE 8.4**  
Properties of Sugar Palm Starch in Comparison With Sago Starch (Adawiyah et al., 2013).

Characterization	Parameters	Sugar Palm Starch	Sago Starch
Chemical composition	Amylose (%w/w)	37.6 ± 1.46	36.6 ± 1.55
	Fat (%w/w)	0.27 ± 0.00	0.24 ± 0.00
	Protein (%w/w)	0.10 ± 0.00	0.08 ± 0.00
	Moisture (%w/w)	9.03 ± 0.00	9.17 ± 0.00
	Ash (%w/w)	0.20 ± 0.00	0.16 ± 0.00
Gelatinization properties	Onset temperature (T <sub>O</sub> ) (°C)	63.0 ± 0.12	58.1 ± 0.28
	Peak temperature (T <sub>P</sub> ) (°C)	67.7 ± 0.07	67.3 ± 0.21
	Conclusion temperature (T <sub>C</sub> ) (°C)	74.6 ± 0.42	79.4 ± 0.88
	Range (T <sub>C</sub> –T <sub>O</sub> ) (°C)	11.6 ± 0.49	21.3 ± 0.79
	ΔH (J/g)	15.4 ± 0.25	16.4 ± 0.24
Mechanical properties	Stress at 10% strain (kPa)	0.61 ± 0.10	0.41 ± 0.04
	Stress at shoulder point (kPa)	23.0 ± 3.65	15.5 ± 0.96
	Strain at shoulder point (%)	54.4 ± 3.54	59.1 ± 2.28
	Working until shoulder point (N mm)	20.2 ± 1.80	14.1 ± 0.82
	Breaking stress (kPa)	29.8 ± 2.64	–
	Breaking strain (%)	60.1 ± 2.61	–
	Work until breaking point (N mm)	29.6 ± 2.45	–
	Compressive force after breaking at 70% strain (N)	8.77 ± 0.59	9.97 ± 1.11
	Compressive force at 90% strain (N)	43.8 ± 2.34	44.0 ± 3.89
	Working until 90% strain (N mm)	108 ± 6.11	90.3 ± 8.37
Adhesive force (N)	–3.64 ± 0.96	–8.99 ± 1.57	

**TABLE 8.5**  
**Mechanical Properties of Starch-Based Biopolymer.**

Film	Tensile Strength (TS, MPa)	Elasticity Modulus (EM, MPa)	Strain at Break ( $\epsilon$ , %)	Year	References
Maize starch	0.24–20	51–315	–	2001	Anglès and Dufresne (2001)
Wheat starch	2.5–7.8	36–301	–	2005	Lu et al. (2005)
Potato starch	3	45	47	2006	Thunwall et al. (2006)
Rice starch	3.2	–	–	2006	Mehyar and Han (2006)
Pea starch	4.2	–	–	2006	Mehyar and Han (2006)
<i>Amaranthus cruentus</i> flour	0.8–3.0	–	74.2–620	2006	Colla et al. (2006)
Pea starch	1.4–5.8	8–98	38–51	2006	Zhang and Han (2006)
Maize starch	1–15	11–320	–	2006	Angellier et al. (2006)
Wheat starch	2.8–6.9	56–480	–	2006	Lu (2006)
Potato starch	13.7	460	–	2007	Kvien et al. (2007)
Corn starch	3	–	20	2008	Dai et al. (2008)
Pea starch	3.9–11.5	31.9–823.9	–	2008	Cao et al. (2008a,b)
Pea starch	3.9–11.9	31.9–498.2	–	2008	Cao et al. (2008a,b)
Maize starch	42	208–838	–	2008	Mathew et al. (2008)
Cassava	1.4–1.6	5–21	30–101	2009	Muller et al. (2009)
Cassava starch	4.8	84.3	–	2009	Teixeira et al. (2009)
Mango puree	8.76	322.05	–	2009	Azeredo et al. (2009)
Maize starch	6.75	220	–	2010	Kaushik et al. (2010)
Pea starch	2.5–12	20.4–210.3	–	2010	Liu et al. (2010)
Wheat starch	3.15–10.98	–	–	2010	Chang et al. (2010)
Corn starch	2.5–3.6	21–533	48–63	2011	Fu et al. (2011)
Rice starch	1.6–11	21–533	3–60	2011	Dias et al. (2011)
Maize starch	0.35	3.12	–	2011	Teixera et al. (2011)
Potato starch	–	460	–	2012	Chen et al. (2012)
Potato starch	17.5	1317.0	–	2013	Hietale et al. (2013)
Potato starch	5.01	160	–	2014	Nasri-Nasrabadi (2014)
Maize starch	2.35	53.6	–	2014	Karimi et al. (2014)
Maize starch	17.4	520	–	2014	Slavutsky and Bertuzzi (2014)
Cush-cush yam starch	1.88	13.9	19	2015	Gutiérrez et al. (2015)
Corn starch	38.0	141.0	–	2015	Babaee et al. (2015)
Potato starch	4.93	–	–	2016	Noshirvani et al. (2016)



Corn starch	11.2	12.4	–	2017	Llanos and Tadini (2018)
Sugar palm starch	4.8	59.97	38.10	2018	Ilyas et al. (2018a)
Yam bean starch	11.47	443	–	2018	Asrofi et al. (2018a)
Sugar palm Starch	11.5	178	–	2018	Ilyas et al. (2018a)
Tapioca starch	5.8	403	–	2018	Asrofi et al. (2018b)
Tapioca starch	12.48	479.8	–	2018	Syafri et al. (2018)
Sugar Palm Starch	10.68	121.26	–	2019	Ilyas et al. (2019c)
Bengkang starch	–	–	–	2019	Syafri et al. (2019)

assume different structures along the polymer chain, based on disulfide cross-link interactions among the amino acid units, hydrophobic, electrostatic, hydrogen bonding, and van der Waals (Fennema, 1985). There are approximately billions of proteins with distinctive properties that can be produced by altering the chain length of polypeptides, the type and ratio of amino acids, and sequence of amino acid. Usually, oilseeds, milk, vegetables, cereals, eggs, and meats (including poultry and fish) have been the main sources of food proteins. According to Fennema (1985), the functional properties of proteins in foods are related to their structural and other physicochemical characteristics. Besides that, the structures of proteins can be altered by various chemical and physical processes such as metal ions, acids and alkalis, lipid interfaces, irradiation, pressure, mechanical treatment, and heat treatment (Fennema, 1985). Such agents have the capacity to change the proteins structures and affect their functional properties. In

films development, these modifications are often used in the formation process to optimize proteins configuration and interactions, resulting in better film properties.

A wide variety of proteins from animal/vegetable sources can be used to produce films, as shown in Table 8.7. In addition to the use of proteins for films production, researchers are focusing on the study of some strategies to improve their performance and to provide bioactive properties. In the group of animal proteins, the most used are caseins, whey protein, collagen, gelatin, myofibrillar proteins, and egg proteins, and among the vegetable proteins, the most used are soy protein, gluten, and zein (Mhd Haniffa et al., 2016; Pérez-Gago and Rhim, 2014).

In addition, casein-based films and biomaterials obtained from caseinate can be found in many applications such as in mulching films, in coatings for vegetables and fruits, in edible films, and in packaging

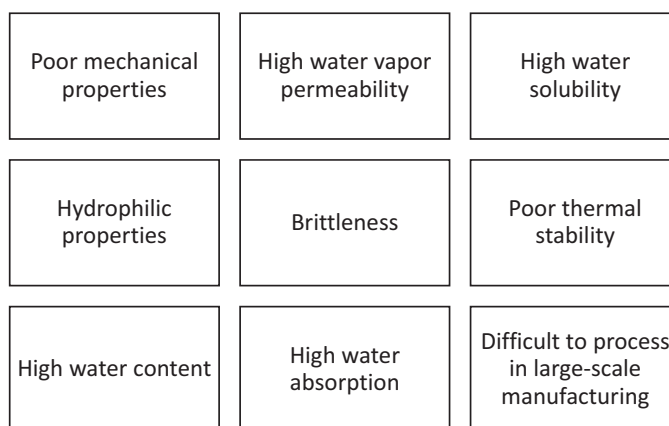


FIG. 8.4 Disadvantages of starch-based biopolymer.

**TABLE 8.6**  
**Starch-Based Biopolymer, Its Manufacturing Technique, and Applications.**

Polymer Component	Manufacturing Techniques	Applications	References
Plasticized starch	Solution casting	Transparent materials	Nasseri and Mohammadi (2014)
Starch	Blending, solution casting	Air permeable, resistant, surface-sized paper, food packaging	Slavutsky and Bertuzzi (2014) and Yang et al. (2014)
Starch	Solution casting	Food packaging	Liu et al. (2010)
Cassava starch	Solution casting	Food packaging	Teixeira et al. (2009)
Sugar palm starch	Solution casting	Food packaging	Ilyas et al. (2018e)
Sugar palm starch	Solution casting	Food packaging	Ilyas et al. (2018b)
Sugar palm starch	Solution casting	Food packaging	Atikah et al. (2019) and Ilyas et al. (2018e)
Wheat starch	Solution casting	Food packaging	Lu et al. (2006)
Tuber native potato	Solution casting	Packaging	Montero et al. (2017)
Cereal corn Starch	Solution casting	Packaging	Montero et al. (2017)
Legume pea starch	Solution casting	Packaging	Montero et al. (2017)
Yam bean	Solution casting	Packaging	Asrofi et al. (2018b)
Yam bean	Solution casting	Packaging	Asrofi et al. (2018a)
Cassava bagasse Starch	Solution casting	Packaging	Teixeira et al. (2009)
Ramie starch	Solution casting	Packaging	Lu et al. (2006)
Potato	Solution casting	Packaging	Chen et al. (2012)
Cassava Starch	Solution casting	Packaging	Syafri et al. (2018)
Bengkuang Starch	Solution casting	Packaging	Syafri et al. (2019)

application. Table 8.8 shows the comparison of mechanical properties of some of the milk protein films formed with plasticizer.

## 5.2 Advantages and Disadvantages of Protein Biopolymer

There are several advantages of protein biopolymer, especially for food packaging application such as highly nutritional quality, good potential to adequately protect food product from their surrounding environment and excellent sensory properties (Gupta and Nayak, 2015). Proteins also act as a flavor and carrier of antioxidant, besides improving the quality of food and bacteriostats. The proteins such as milk proteins (whey proteins and casein) (Su et al., 2010), gluten (Zhong and Yuan, 2013), corn (Aydt et al., 1991), peanut (Aydt et al., 1991), sunflower seed (Martinez et al., 2005), whey protein (Jooyandeh, 2011), soy bean

(Zhang et al., 2010), gelatin from collagen (Gómez-Guillén et al., 2011), soy protein (Tian et al., 2011), and wheat (Aydt et al., 1991) are suitable for the fabrication of protein biopolymer film due to its nutritional properties.

This type of biopolymer is also being used in nonedible packaging. Protein-based biopolymer also had impressed gas barrier properties compared with those prepared from polysaccharides and lipids (Cuq et al., 1998). Amazingly, the oxygen gas (O<sub>2</sub>) permeability of the soy protein film was 670, 540, 500, and 260 times lower than that of pectin, starch, low density PE, and methyl cellulose, respectively, when they are not moist (Cuq et al., 1998). One of the disadvantage of soy protein film is low moisture barrier properties. This is because of their hydrophilic property and the considerable amount of hydrophilic plasticizer used in film preparation (Cuq et al., 1998).

TABLE 8.7

Some Research Using Different Protein Bases in the Preparation of Films Combining Strategies to Improve the Properties of Films.

Protein Type	Strategies	Effect Observed	References
<b>VEGETABLE PROTEIN</b>			
Amaranth	Native waxy and maize starch nanocrystals	Structure reinforcement	Condés et al. (2015)
Bitter vetch ( <i>Vicia ervilia</i> ) seed/corn zein	Bilayer	Improved structure	Arabestani et al. (2016)
Canola	—	—	Shi and Dumont (2014)
Pea	Lysozyme	Antimicrobial	Fabra et al. (2014)
Sesame meal	—	—	Sharma and Singh (2016)
Soy	Chestnut ( <i>Castanea mollissima</i> ) bur extracts	Antioxidant	Wang et al. (2016)
Soy protein isolate	Peanut protein nanoparticles	Structure reinforcement	Li et al. (2015)
Soy/agar	Blend/extrusion	Structure reinforcement	Garrido et al. (2016)
Sunflower protein	Clove essential oil	Structure reinforcement and improved food shelf life	Salgado et al. (2013)
<b>ANIMAL PROTEIN</b>			
Argentine anchovy ( <i>Engraulis anchoita</i> )	Sorbic or benzoic acids	Antimicrobial	Rocha et al. (2014)
Chicken feet	—	—	Lee et al. (2015a)
Fish gelatin	Chitosan nanoparticles	Structure reinforcement	Hosseini et al. (2015)
Gelatin	Longan seed extract	Antioxidant	Sai-Ut et al. (2015)
Porcine meat and bone meal	Coriander oil	Antimicrobial	Lee et al. (2015b)
Shrimp ( <i>Litopenaeus vannamei</i> ) muscle	Cinnamaldehyde/thermal treatment	Cross-linking	Gómez-Estaca et al. (2014)
Whey protein	Gum tragacanth	Structure reinforcement	Tonyali et al. (2018)
Whey protein	Ultraviolet treatment	Structure reinforcement	Díaz et al. (2017)
Whey protein isolate	<i>Lactobacillus rhamnosus</i>	Antimicrobial	Beristain-Bauza et al. (2016)
Whitemouth croaker muscle	Pink pepper phenolic	Browning reduction	Romani et al. (2018)

### 5.3 Application of Protein Biopolymer

Several scientists had conducted studied on the protein-based edible films such as corn zein films on nut and fruit products, whey protein film, casein emulsion film, and soy protein film for food packaging applications (Calva-Estrada et al., 2019; Gennadios, 2004; Rhim et al., 2004; Schmid and Müller, 2019). The polymeric characteristics of the protein film have been used for edible food packaging application (Khwaldia et al., 2010; Oussalah et al., 2004; Su et al., 2010; Zhang et al., 2010), but for nonfood packaging application, the

major problems are an advancement of mechanical properties (such as tensile modulus, shear strength, flexural, elasticity, strength, and toughness). Table 8.9 displays packaging and biomedical applications of protein biopolymer. Besides that, biocompatible materials from proteins biomaterials have been utilized to develop scaffolds for various biomedical applications such as drug delivery, tissue engineering, wound dressings, and membrane filters. Fig. 8.5 shows the images of CMC (carboxymethyl cellulose)/SPI (soy protein isolate) film. This film is made up by a continuous

**TABLE 8.8**  
Mechanical Properties of the Milk Protein Film Formed in the Presence of Different Formulation of Plasticizers.

Film	Tensile Strength (MPa)	Elongation at Break (%)	References
Sodium caseinate/glycerol (4:1)	10.5	17.4–26.7	Siew et al. (1999)
Sodium caseinate/glycerol (2:1)	73.7–84.2	10.9–11.7	
Sodium caseinate/PEG (4.54:1)	5.3	10.9–16.35	
Sodium caseinate/PEG (1.9:1)	25.4	10.9–13.9	
Whey protein/glycerol (5.7/1)	4.1	29.1	McHugh et al. (1994) and McHugh and Krochta (1994)
Whey protein/glycerol (2.3/1)	30.8	13.9	
Whey protein/sorbitol (2.3/1)	1.6	14.0	
Whey protein/sorbitol (1/1)	8.7	14.7	

casting method. Remarkably, this film can be produced simply with absence of puncture and cracks. Besides that, the film produced has a good flexible and durability, as much as it is necessary to be rolled into forms for sensible applications (Su et al., 2010).

## 6 CHITIN AND CHITOSAN

### 6.1 Properties of Chitin and Chitosan

Chitin is a polysaccharide with linear chains comprising of 2-acetamide-2-deoxy- $\beta$ -D-glucopyranose units, which

**TABLE 8.9**  
Packaging and Biomedical Applications Made From Protein Biopolymer.

Protein Biopolymer	References
<b>PACKAGING APPLICATION</b>	
Gluten films	Hemsri et al. (2011), Zhong and Yuan (2013), Zuo et al. (2009)
Milk protein films	Banerjee et al. (1996), Chen (1995), Kinsella and Morr (1984), Su et al. (2010), Vachon et al. (2000), Yoo and Krochta (2011)
Soy protein films	Park et al. (2000), Rangavajhyala et al. (1997), Rhim et al. (2006), Stuchell and Krochta (1994), Su et al. (2010), Tian et al. (2011), Zhang et al. (2010)
Corn zein films	Khwaldia et al. (2010), Lim and Jane (1994), Pol et al. (2002)
Gelatin films	Avena-Bustillos et al. (2011), Carvalho and Grosso (2006), Sobral et al. (2001)
Silk protein films	Jiang et al. (2007), Zhang et al. (2004)
<b>BIOMEDICAL APPLICATION</b>	
Scaffold in tissue engineering	Haghpanah et al. (2009), Hu et al. (2010)
Drug delivery systems	Cho et al. (2008), Koutsopoulos et al. (2009), Qiu and Park (2001), Torchilin (2005)
Biosynthetic hybrid hydrogels scaffold	Almany and Seliktar (2005)

are linked by glycosidic bonds  $\beta(1 \rightarrow 4)$  (Ma et al., 2019). Chitin exists as the second most abundant organic substance in the biosphere, surpassed only by cellulose, but chitin surpasses the latter in terms of replacement rate, which is twice as high as cellulose (Deringer et al., 2016). Chitin is found in the skeletal structure of invertebrates, such as arthropods, annelids, mollusks, and coelenterates, and in the cell walls of



**FIG. 8.5** Photographs of CMC/SPI film in expanded and rolled states fabricated by a continuous casting method (Su et al., 2010).

diatoms and some fungi (Nataraj et al., 2018). Depending on the organism, chitin adopts different polymorphic structures called  $\alpha$ -,  $\beta$ - and  $\gamma$ -chitin. The different polymorphic structures of chitin correspond to different arrangements in solid state (Pighinelli, 2019). The  $\alpha$ -chitin is found in rigid and resistant structures, such as the arthropod cuticle. This chitin is usually associated with proteins, inorganic materials, or both (Deringer et al., 2016). The  $\beta$ - and  $\gamma$ -chitin occur in flexible structures and are also resistant. The  $\alpha$ -chitin is the most abundant form and is considered the most stable, considering that the conversion of  $\beta$ - and  $\gamma$ -chitin in the first one is irreversible (Deringer et al., 2016).

Chitosan is commonly made by alkaline deacetylation of  $\alpha$ -chitin. Chitin and chitosan are important linear polysaccharides consisting, respectively, of 2-acetamide-2-deoxy- $\beta$ -D-glucopyranose (GlcNAc) and 2-amino-2-deoxy- $\beta$ -D-glucopyranose (GlcN) linked by  $\beta(1 \rightarrow 4)$  (Nataraj et al., 2018). Fig. 8.6 shows the molecular structures of chitin and chitosan. Chitosan solubilization can be attributed to protonation of the  $-\text{NH}_2$  in the C-2 of D-glucosamine units in acid medium, which results in the conversion into a polyelectrolyte

(Roy et al., 2017). The polycationic character of chitosan in acid medium is due to its weak base characteristic, thus its amino groups are easily protonated (Nataraj et al., 2018). Moreover, the hydroxyl groups of carbons 3 and 6 can also be protonated, which increases chitosan reactivity (Nataraj et al., 2018).

## 6.2 Advantages and Disadvantages of Chitin and Chitosan

Chitin is also fascinating in cosmetology because it is tolerable by the skin. It acts as an effective hydrating agent and a film-forming tensor having two benefits that are often cited: it provides water and it minimizes dehydration. The molecular weight of most chitosan-based products is too high that they cannot penetrate the skin, which is a crucial characteristic of a skincare product. These materials consist of chitosan hydrochloride, chitosan acetate, chitosan lactate, carboxymethyl chitosan, quaternized derivatives, oligosaccharides, and chitin sulfate and carboxymethyl chitin (Pighinelli, 2019).

Chitosan films have revealed remarkable results, possessing good mechanical properties and having the

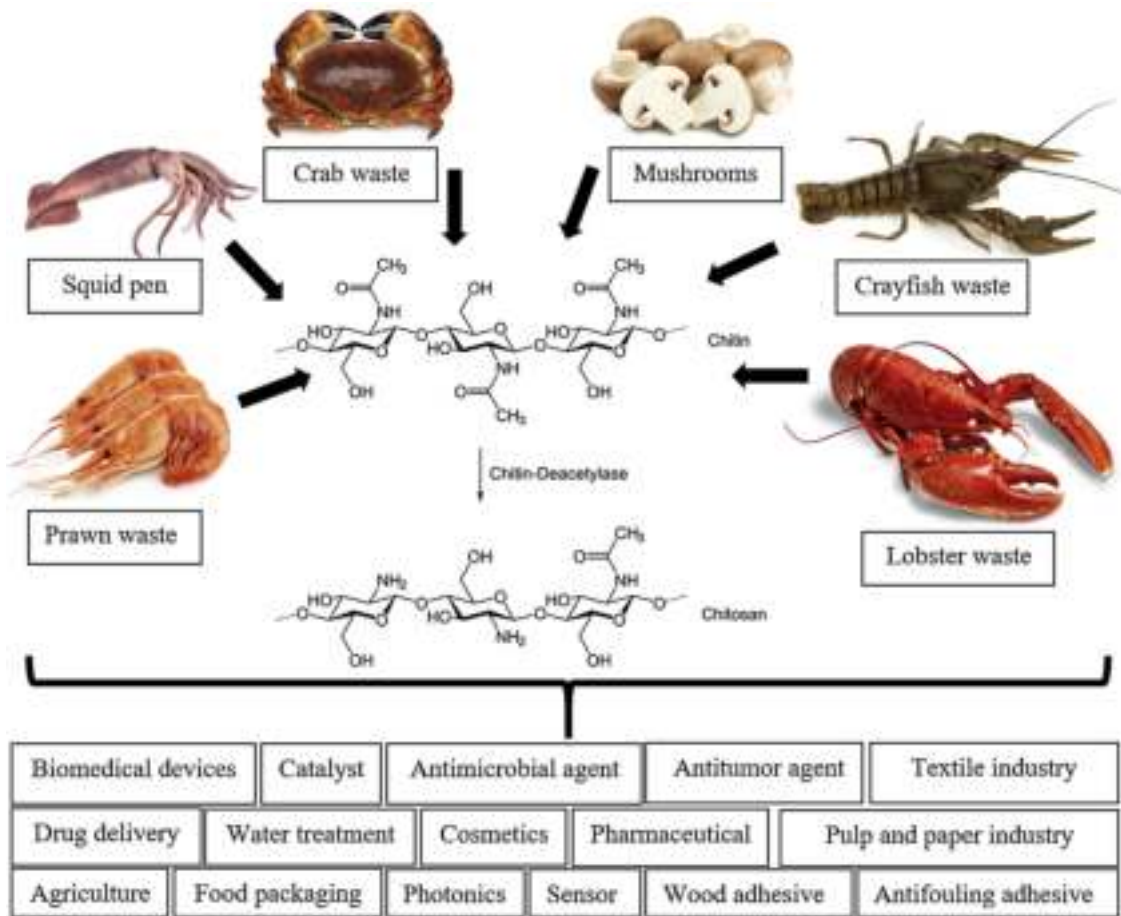


FIG. 8.6 Molecular structures of chitin and chitosan, their sources and applications.

advantage of the ability to integrate functional substances, for instance, as vitamins and carriers that release antimicrobial agents. Chitosan derivatives are advantageous due to the biocompatibility and safe, nontoxic to living tissues and their hydrophilic property, biodegradability, antibacterial activity, bioadhesivity, mucoadhesivity, and complexing property (Llanos and Tadini, 2018).

Chitosan has the ability to form gels in addition to having viscosity-related properties, complete biodegradability, and even antitumor influence, like alginate polysaccharide. Chitosan creates films permeable to air that aid cellular regeneration while protecting tissues from microbial attacks. Chitosan also stimulates the process of regeneration of tissues, making it a suitable material in artificial skin manufacturing for the applications in skin grafts on high degree burns and in surgical applications (suture threads) (Oryan and Sahvieh,

2017). In terms of health benefit, chitosan is able to trap lipids at its insolubilization pH along the digestive tract, which significantly reduces cholesterol level in the blood. Chitosan possesses bioadhesive properties that make it of interest in adhesive-sustained release formulation required. Mucoadhesivity permits to enhance the adsorption of drugs especially at neutral pH.

This natural polymer owns several inherent features making it an effective material for environmental purposes: (i) lower cost in comparison with activated carbon or organic resins available in market, (ii) outstanding pollutant-adhering capacities and outstanding selectivity, (iii) versatility, and (iv) biodegradability. Indeed, major applications of chitosan are based on its excellent capability to tightly bind a whole range of pollutants (Vidal and Moraes, 2019).

Researchers had summarized that the biosorbents are effective in pollutant elimination with the extra

advantage of being cheap, nontoxic, and biocompatible.

Chitosan has gained substantial attention for developing microcapsules, which is advantageous as drug carriers due to their controlled release properties and biocompatibility (Rokhade et al., 2007). Presence of microcapsules also makes chitosan applicable for wall materials for textile finishing product encapsulation (Alonso et al., 2010). Numerous techniques have been used in the creation of chitosan microcapsules, such as spray drying and phase coacervation (Liu et al., 2011); the microcapsules yield is either single or multilayer, depending on microencapsulation method (Pothakamury and Barbosa-Cánovas, 1995).

Chitin and chitosan poses high organic solvent resistance, which is useful for separation membranes. This biomaterial is used with organic solvents, where chemical resistance is typical. Explicitly, chitin is highly acid resistant, meanwhile chitosan is highly alkaline resistant. These features contribute to the application of chitin and chitosan as separation membranes for a range of uses in response to specific requirements (Chaudhary et al., 2015).

However, chitosan films are unsuitable for packaging application due to the fact that they are highly permeable to water vapor. In addition, their hydrophilic

character also make them to exhibit resistance to fat diffusion and selective gas permeability (Morin-Crini et al., 2019).

### 6.3 Application of Chitin and Chitosan

Chitin- or chitosan-based biomaterials are promising candidates to be used for wound healing, tissue engineering, and drug delivery. The fact that they are originated from the freely available natural sources has made them more economically stable compared with synthetic polymer materials. Moreover, their utilizations in biomedical field, such as scaffolds for tissue engineering, manages to minimize cost and manpower needed for second surgery to remove them, since chitin/chitosan-based materials are biodegradable. Besides, their biocompatibility also avoids the necessity for any treatments to be performed due to the rejection of implants fabricated from employing these materials. Currently existing chitin- and chitosan-based commercial products are as summarized in Table 8.10. The markets for chitin and chitosan are the United States, China, Norway, France, Poland, Japan, Germany, Korea, Canada, Australia, and the United Kingdom (Crini and Lichtfouse, 2016; Morin-Crini et al., 2019). In 2015, Japan has dominated (advanced in technology, commercialization, and utilization of these

**TABLE 8.10**  
Commercial Products From Chitin and Chitosan and Their Applications.

Product	Manufacturer	Application	Patent/References
LipoSan Ultra	Primex	Weight loss	US 6130321, Johnson and Nichols (2000)
Slim MED	KitoZyme	Weight management and treatment	US 20040126444 A1, D'huart and Dallas (2004)
ChitoDot	Tricol biomedical, Inc.	Wound dressing and bleeding control	US 8269058 B2, McCarthy et al. (2012)
BST-Gel	Piramal Healthcare (Canada) Inc.	Chronic wound healing, bone filling, cartilage repair, intervertebral disc regeneration	US 8747899, Chaput and Chenite (2014)
ChitoFlex PRO	Tricol biomedical, Inc.	Wound dressing and bleeding control	US 8668924 B2, McCarthy et al. (2014)
HemCon ChitoGauze PRO	Tricol Biomedical, Inc.	Wound dressing material	US 9205170 B2, Lucchesi and Xie (2015)
Talymed	Marine Polymer Technologies	Wound dressing material	US 9139664 B2, Finkielstein and Vournakis (2015)
Protasan	NovaMatrix	Pharmaceutical application	WO 2015081304 A1, Francis et al. (2015)
Reaxon	Medovent	Nerve rejuvenation	

biopolymers) the industry market of chitin and chitosan accounting for 35%, which is equivalent to 700–800 tons per annual (Morin-Crini et al., 2019).

## 7 POLY(BUTYLENE SUCCINATE)

### 7.1 Properties of PBS

The world community is anticipated to grow to 9 billion by 2050, hence resulting in the increment of the plastic production and undoubtedly, plastic wastes (Emadian et al., 2017). These motivated researchers to explore and introduce bioplastics, such as PCL, polylactide (PLA), PBS, and TPS. PBS is identified as the most notable biopolymer that is produced via polycondensation of butanediol and succinic acid having unique features, such as good melt processability, great toughness, high chemical resistance, high heat distortion temperature, biodegradable, good mechanical properties, high chemical, and thermal resistance (Boonprasith et al., 2013). This biopolymer is derived from natural sources (Jamaluddin et al., 2016). PBS is a thermoplastic polymer resin in the polyester family. PBS comprises of polymerized units of butylene succinate with repeating  $C_8H_{12}O_4$  units. There are two ways to synthesize PBS:

1. Transesterification process (from succinate diesters)
2. Direct esterification process (from the diacid).

The typical route in PBS production is direct esterification of succinic acid with 1,4-butanediol, which involves two steps. First, excess of diol is esterified with diacid to form PBS oligomers with water as by-product of process. Next, these PBS oligomers are transesterified under vacuum condition with the presence of catalyst (zirconium, germanium, titanium, or tin) in order to produce a high molecular weight (Mw) polymer that is PBS as shown in Fig. 8.7.

The physical properties of poly(butylene succinate-co-butylene adipate) copolyesters can vary with comonomer content, as tabulated in Table 8.11 and Fig. 8.8.

### 7.2 Advantages and Disadvantages of PBS

PBS are synthesized using succinic acid. Succinic acid is also obtainable from the fermentation of microorganisms on renewable feedstocks, for instance, glucose, starch, xylose, etc (Song and Lee, 2006). *Actinobacillus succinogenes*, *Anaerobiospirillum succiniciproducens*, *Mannheimia succiniciproducens*, and recombinant *Escherichia coli* are well-known and well-established bacterial production strains that can produce succinic acid (Gigli et al., 2016; Song and Lee, 2006; Xu and Guo, 2010).

Biodegradability is another useful characteristic of PBS biopolymer, since the need of surgery to remove the carrier/implant can be prevented as it self-degrades when its desired function has ended. It is worth stating that not only the biomaterial but also the degradation products are nonhazardous for the host.

Conversely, PBS has attention-grabbing physiochemical properties and can be simply synthesized by melt polycondensation at reasonable costs. Added value is given by the ability to achieve both succinic acid and 1,4-butanediol from renewable resources, which marks PBS a completely bio-based and biodegradable polymer.

### 7.3 Application of PBS

As PBS disintegrates naturally into water and  $CO_2$ , it offers as a biodegradable substitute to some commonly used plastics. The range of PBS application areas is still expanding and several areas can be recognized, but it remains hard to know explicitly in which specific object PBS is really used. In packaging field, PBS is converted into films, bags, or boxes, for both food and cosmetic packaging. PBS also could be found in disposable merchandises, such as tableware or medical articles. Next, in agriculture, PBS is useful in the manufacturing of mulching films or delayed release materials for pesticide and fertilizer. PBS is also promising to find market shares in fishery (for fishing nets), forestry, civil

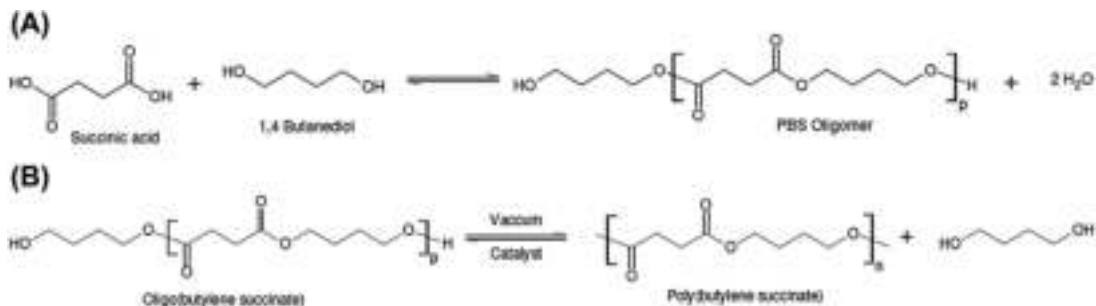


FIG. 8.7 PBS synthetization process using direct esterification process: (A) first step and (B) second step (Jamaluddin et al., 2016).



**TABLE 8.11**  
Thermal Properties and Degree of Crystallinity of PBSA Random Polyesters (Xu and Guo, 2010).

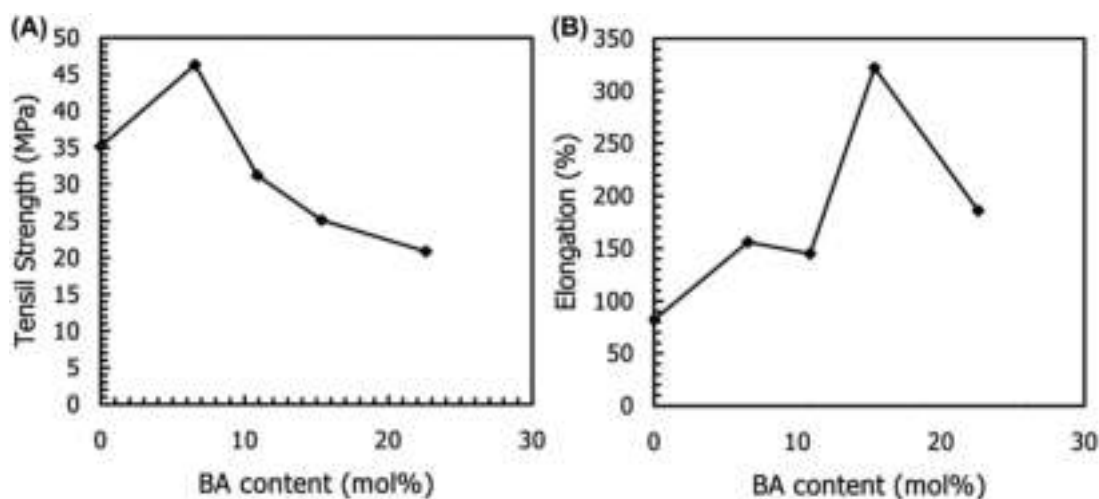
Polymer	$\Delta H_m$ (J/g)	$\Delta H_m^c$ (J/g)	$T_m$ (°C)	$T_g$ (°C) <sup>a</sup>	Crystallinity <sup>b</sup> (%)	Crystallinity <sup>c</sup> (%)
PBS	67.4	110.3	112	-18	61.1	39.66
PBSA-5 <sup>d</sup>	96.0	110.3	108	-21	87.0	54.47
PBSA-10	72.5	110.3	103	-23	65.7	45.83
PBSA-15	79.8	110.3	99	-27	72.3	45.27
PBSA-20	59.5	110.3	92	-34	53.9	46.83

<sup>a</sup> The glass transition temperature ( $T_g$ ) was adopted from the  $\tan\delta$  peak measured by dynamic thermal analysis.

<sup>b</sup> The degree of crystallinity was the ratio of melting enthalpy determined by DSC to the melting enthalpy of completely crystalline PBS (100 J/g).

<sup>c</sup> The degree of crystalline was calculated from WAXD results.

<sup>d</sup> The number indicates the molar percentage of adipic acid in the total feed acids for synthesis of PBSA copolyester.



**FIG. 8.8** The mechanical properties of PBSA at different contents of butylene adipate (BA) content: **(A)** Tensile strength and **(B)** Elongation at break (Xu and Guo, 2010).

engineering, or other fields in which recovery and recycling of materials after utilization is challenging. In medical field, PBS is adopted as biodegradable drug encapsulation system and is also being studied for implants.

## 8 SUMMARY AND FUTURE PERSPECTIVES

The progress of bio-based polymers as substitution for petroleum-based synthetics has been an area of attention due to the nondegradable and nonrenewable

nature of synthetic plastics, as well as a primary research challenge for scientists. With the future fossil fuel crisis, the exploration and expansion of alternative chemical/material alternatives is crucial in minimizing mankind's reliance on fossil fuel resources. Some of the probable substitute candidates are PHA, PLA, starch, protein, chitin, chitosan, and PBS. Bio-based polymers are generally defined as polymers manufactured from renewable resources, comprising of three different categories: (1) natural polymers originated from biomass such as the agropolymers from agrosources, e.g.,

starch, cellulose, protein, chitin, and chitosan; (2) polymers formed by microorganisms, e.g., PHA; and (3) synthetic biopolymers, which are chemically synthesized using monomers obtained from agroresources, e.g., poly(lactic acid) and PBS, which is biodegradable as well. Overall, bio-based polymers are still new, though, they are in continuous development. Research attempts are obviously intent on addressing challenges constraining the use of bio-based polymers, comprising reduction the production and processing costs, improving barrier and mechanical properties, or introducing extra functions as active and smart packaging. Being a carbon neutral and valuable polymer manufactured from many renewable carbon sources by microorganisms, PHA is said to be a sustainable and environmental-friendly material. However, nowadays, PHA is not cost competitive compared with fossil-derived products. PLA is also produced from renewable sources having high tensile strength and modulus and can be processed via conventional processing methods. Starch complies all the principle aspects; hence, it is suitable for edible coatings/films. Chitin is the most abundant natural amino polysaccharide and is predicted to be produced yearly almost as much as cellulose. It has grabbed great attention not only as an underutilized resource but also as a new useful material of great potential in many fields, and current progress in chitin chemistry is quite remarkable. PBS is a renowned aliphatic polyester, provided its fascinating thermomechanical properties and the proven biodegradability, combined with acceptable raw material and production costs. As a conclusion, biodegradable plastics are often reflected as savior products. Shifting to these materials would lessen carbon dioxide, greenhouse gas emission levels, energy of manufacturing, and amount of waste produced and create opportunity for new plastic industries.

## REFERENCES

- Abral, H., Ariksa, J., Mahardika, M., Handayani, D., Aminah, I., Sandrawati, N., Pratama, A.B., Fajri, N., Sapuan, S.M., Ilyas, R.A., 2020a. Transparent and antimicrobial cellulose film from ginger nanofiber. *Food Hydrocolloids* 98, 105266. <https://doi.org/10.1016/j.foodhyd.2019.105266>.
- Abral, H., Ariksa, J., Mahardika, M., Handayani, D., Aminah, I., Sandrawati, N., Sapuan, S.M., Ilyas, R.A., 2019a. Highly transparent and antimicrobial PVA based bionanocomposites reinforced by ginger nanofiber. *Polymer Testing* 106186. <https://doi.org/10.1016/j.polymertesting.2019.106186>.
- Abral, H., Atmajaya, A., Mahardika, M., Hafizulhaq, F., Kadriadi, Handayani, D., Sapuan, S.M., Ilyas, R.A., 2020b. Effect of ultrasonication duration of polyvinyl alcohol (PVA) gel on characterizations of PVA film. *Journal of Materials Research and Technology* 1–10. <https://doi.org/10.1016/j.jmrt.2019.12.078>.
- Abral, H., Basri, A., Muhammad, F., Fernando, Y., Hafizulhaq, F., Mahardika, M., Sugiarti, E., Sapuan, S.M., Ilyas, R.A., Stephane, I., 2019b. A simple method for improving the properties of the sago starch films prepared by using ultrasonication treatment. *Food Hydrocolloids* 93, 276–283. <https://doi.org/10.1016/j.foodhyd.2019.02.012>.
- Adawiyah, D.R., Sasaki, T., Kohyama, K., 2013. Characterization of arenga starch in comparison with sago starch. *Carbohydrate Polymers* 92, 2306–2313. <https://doi.org/10.1016/j.carbpol.2012.12.014>.
- Aisyah, H.A., Paridah, M.T., Sapuan, S.M., Khalina, A., Berkalp, O.B., Lee, S.H., Lee, C.H., Nurazzi, N.M., Ramli, N., Wahab, M.S., Ilyas, R.A., 2019. Thermal properties of woven kenaf/carbon fibre-reinforced epoxy hybrid composite panels. *International Journal of Polymer Science* 1–8. <https://doi.org/10.1155/2019/5258621>.
- Almany, L., Seliktar, D., 2005. Biosynthetic hydrogel scaffolds made from fibrinogen and polyethylene glycol for 3D cell cultures. *Biomaterials* 26, 2467–2477. <https://doi.org/10.1016/j.biomaterials.2004.06.047>.
- Alonso, D., Gimeno, M., Sepúlveda-Sánchez, J.D., Shirai, K., 2010. Chitosan-based microcapsules containing grapefruit seed extract grafted onto cellulose fibers by a non-toxic procedure. *Carbohydrate Research* 345, 854–859. <https://doi.org/10.1016/j.carres.2010.01.018>.
- Angellier, H., Molina-Boisseau, S., Dole, P., Dufresne, A., 2006. Thermoplastic starch–waxy maize starch nanocrystals nanocomposites. *Biomacromolecules* 7, 531–539. <https://doi.org/10.1021/bm050797s>.
- Anglès, M.N., Dufresne, A., 2001. Plasticized starch/tunicin whiskers nanocomposite materials. 2. Mechanical behavior. *Macromolecules* 34, 2921–2931. <https://doi.org/10.1021/ma001555h>.
- Arabestani, A., Kadivar, M., Amoresano, A., Illiano, A., Di Piero, P., Porta, R., 2016. Bitter vetch (*Vicia ervilia*) seed protein concentrate as possible source for production of bilayered films and biodegradable containers. *Food Hydrocolloids* 60, 232–242. <https://doi.org/10.1016/j.foodhyd.2016.03.029>.
- Asrofi, M., Abral, H., Kasim, A., Pratoto, A., Mahardika, M., Hafizulhaq, F., 2018a. Mechanical properties of a water hyacinth nanofiber cellulose reinforced thermoplastic starch bionanocomposite: effect of ultrasonic vibration during processing. *Fibers* 6, 40. <https://doi.org/10.3390/fib6020040>.
- Asrofi, M., Abral, H., Kasim, A., Pratoto, A., Mahardika, M., Hafizulhaq, F., 2018b. Characterization of the sonicated yam bean starch bionanocomposites reinforced by nanocellulose water hyacinth fiber (WHF): the effect of various fiber loading. *Journal of Engineering Science and Technology* 13, 2700–2715.
- Asyraf, M.R.M., Ishak, M.R., Sapuan, S.M., Yidris, N., 2020. Woods and composites cantilever beam: a comprehensive review of experimental and numerical creep

- methodologies. *Journal of Materials Research and Technology*. <https://doi.org/10.1016/j.jmrt.2020.01.0>.
- Atikah, M.S.N., Ilyas, R.A., Sapuan, S.M., Ishak, M.R., Zainudin, E.S., Ibrahim, R., Atiqah, A., Ansari, M.N.M., Jumaidin, R., 2019. Degradation and physical properties of sugar palm starch/sugar palm nanofibrillated cellulose bionanocomposite. *Polimery* 64, 27–36. <https://doi.org/10.14314/polimery.2019.10.5>.
- Atiqah, A., Jawaid, M., Sapuan, S.M., Ishak, M.R., Ansari, M.N.M., Ilyas, R.A., 2019. Physical and thermal properties of treated sugar palm/glass fibre reinforced thermoplastic polyurethane hybrid composites. *Journal of Materials Research and Technology* 8, 3726–3732. <https://doi.org/10.1016/j.jmrt.2019.06.032>.
- Auras, R.A., Singh, S.P., Singh, J.J., 2005. Evaluation of oriented poly(lactide) polymers vs. existing PET and oriented PS for fresh food service containers. *Packaging Technology and Science* 18, 207–216. <https://doi.org/10.1002/pts.692>.
- Avena-Bustillos, R.J., Chiou, B., Olsen, C.W., Bechtel, P.J., Olson, D.A., McHugh, T.H., 2011. Gelation, oxygen permeability, and mechanical properties of mammalian and fish gelatin films. *Journal of Food Science* 76, E519–E524. <https://doi.org/10.1111/j.1750-3841.2011.02312.x>.
- Aydt, T.P., Weller, C.L., Testin, R.F., 1991. Mechanical and barrier properties of edible corn and wheat protein films. *Transactions of the American Society of Agricultural Engineers* 34, 207–211. <https://doi.org/10.13031/2013.31646>.
- Azammi, A.M.N., Ilyas, R.A., Sapuan, S.M., Ibrahim, R., Atikah, M.S.N., Asrofi, M., Atiqah, A., 2020. Characterization studies of biopolymeric matrix and cellulose fibres based composites related to functionalized fibre-matrix interface. In: *Interfaces in Particle and Fibre Reinforced Composites*. Elsevier, London, pp. 29–93. <https://doi.org/10.1016/B978-0-08-102665-6.00003-0>.
- Azeredo, H.M.C., Mattoso, L.H.C., Wood, D., Williams, T.G., Avena-Bustillos, R.J., McHugh, T.H., 2009. Nanocomposite edible films from mango puree reinforced with cellulose nanofibers. *Journal of Food Science* 74. <https://doi.org/10.1111/j.1750-3841.2009.01186.x>.
- Babae, M., Jonoobi, M., Hamzeh, Y., Ashori, A., 2015. Biodegradability and mechanical properties of reinforced starch nanocomposites using cellulose nanofibers. *Carbohydrate Polymers* 132, 1–8. <https://doi.org/10.1016/j.carbpol.2015.06.043>.
- Banerjee, R., Chen, H., Wu, J., 1996. Milk protein-based edible film mechanical strength changes due to ultrasound process. *Journal of Food Science* 61, 824–828. <https://doi.org/10.1111/j.1365-2621.1996.tb12211.x>.
- Beristain-Bauza, S.C., Mani-López, E., Palou, E., López-Malo, A., 2016. Antimicrobial activity and physical properties of protein films added with cell-free supernatant of *Lactobacillus rhamnosus*. *Food Control* 62, 44–51. <https://doi.org/10.1016/j.foodcont.2015.10.007>.
- Boonprasith, P., Wootthikanokkhan, J., Nimitsiriwat, N., 2013. Mechanical, thermal, and barrier properties of nanocomposites based on poly(butylene succinate)/thermoplastic starch blends containing different types of clay. *Journal of Applied Polymer Science* 130, 1114–1123. <https://doi.org/10.1002/app.39281>.
- Calva-Estrada, S.J., Jiménez-Fernández, M., Lugo-Cervantes, E., 2019. Protein-based films: advances in the development of biomaterials applicable to food packaging. *Food Engineering Reviews* 11, 78–92. <https://doi.org/10.1007/s12393-019-09189-w>.
- Cao, X., Chen, Y., Chang, P.R., Muir, A.D., Falk, G., 2008a. Starch-based nanocomposites reinforced with flax cellulose nanocrystals. *Express Polymer Letters* 2, 502–510. <https://doi.org/10.3144/expresspolymlett.2008.60>.
- Cao, X., Chen, Y., Chang, P.R., Stumborg, M., Huneault, M.A., 2008b. Green composites reinforced with hemp nanocrystals in plasticized starch. *Journal of Applied Polymer Science* 109, 3804–3810. <https://doi.org/10.1002/app.28418>.
- Carvalho, R.A. de, Grosso, C.R.F., 2006. Properties of chemically modified gelatin films. *Brazilian Journal of Chemical Engineering* 23, 45–53. <https://doi.org/10.1590/S0104-66322006000100006>.
- Chang, P.R., Jian, R., Zheng, P., Yu, J., Ma, X., 2010. Preparation and properties of glycerol plasticized-starch (GPS)/cellulose nanoparticle (CN) composites. *Carbohydrate Polymers* 79, 301–305. <https://doi.org/10.1016/j.carbpol.2009.08.007>.
- Chaput, C., Chenite, A., June 10, 2014. Injectable in situ self-forming mineral-polymer hybrid composition and uses thereof. US Patent 8747899.
- Chaudhary, J.P., Vadodariya, N., Nataraj, S.K., Meena, R., 2015. Chitosan-based aerogel membrane for robust oil-in-water emulsion separation. *ACS Applied Materials & Interfaces* 7, 24957–24962. <https://doi.org/10.1021/acsami.5b08705>.
- Chen, D., Lawton, D., Thompson, M.R., Liu, Q., 2012. Biocomposites reinforced with cellulose nanocrystals derived from potato peel waste. *Carbohydrate Polymers* 90, 709–716. <https://doi.org/10.1016/j.carbpol.2012.06.002>.
- Chen, G.-Q., 2010. Nodax™ class PHA copolymers: their properties and applications - industrial production of PHA. *Plastics From Bacteria: Natural Functions and Applications* 14, 121–132. <https://doi.org/10.1007/978-3-642-03287>.
- Chen, H., 1995. Functional properties and applications of edible films made of milk proteins. *Journal of Dairy Science* 78, 2563–2583. [https://doi.org/10.3168/jds.S0022-0302\(95\)76885-0](https://doi.org/10.3168/jds.S0022-0302(95)76885-0).
- Cho, K., Wang, X., Nie, S., Chen, Z., Shin, D.M., 2008. Therapeutic nanoparticles for drug delivery in cancer. *Clinical Cancer Research* 14, 1310–1316. <https://doi.org/10.1158/1078-0432.CCR-07-1441>.
- Colla, E., do Amaral Sobral, P.J., Menegalli, F.C., 2006. *Amaranthus cruentus* flour edible films: influence of stearic acid addition, plasticizer concentration, and emulsion stirring speed on water vapor permeability and mechanical properties. *Journal of Agricultural and Food Chemistry* 54, 6645–6653. <https://doi.org/10.1021/jf0611217>.
- Condés, M.C., Añón, M.C., Mauri, A.N., Dufresne, A., 2015. Amaranth protein films reinforced with maize starch nanocrystals. *Food Hydrocolloids* 47, 146–157. <https://doi.org/10.1016/j.foodhyd.2015.01.026>.

- Crini, G., Lichtfouse, E., 2016. Sustainable Agriculture Reviews, Sustainable Agriculture Reviews, Sustainable Agriculture Reviews. Springer International Publishing, Cham. <https://doi.org/10.1007/978-3-319-26777-7>.
- Cuq, B., Gontard, N., Guilbert, S., 1998. Proteins as agricultural polymers for packaging production. *Cereal Chemistry Journal* 75, 1–9. <https://doi.org/10.1094/CCHEM.1998.75.1.1>.
- D'huart, J., Dallas, C., July 1, 2004. Cactaceae-based formulation having the property of fixing fats, and method for obtaining same. US Patent 2004/0126444 A1.
- Dai, H., Chang, P.R., Yu, J., Ma, X., 2008. N,N-Bis(2-hydroxyethyl)formamide as a new plasticizer for thermoplastic starch. *Starch - Stärke* 60, 676–684. <https://doi.org/10.1002/star.200800017>.
- Deringer, V.L., Englert, U., Dronskowski, R., 2016. Nature, strength, and cooperativity of the hydrogen-bonding network in  $\alpha$ -chitin. *Biomacromolecules* 17, 996–1003. <https://doi.org/10.1021/acs.biomac.5b01653>.
- Dias, A.B., Müller, C.M.O., Larotonda, F.D.S., Laurindo, J.B., 2011. Mechanical and barrier properties of composite films based on rice flour and cellulose fibers. *Lebensmittel-Wissenschaft und -Technologie- Food Science and Technology* 44, 535–542. <https://doi.org/10.1016/j.lwt.2010.07.006>.
- Díaz, O., Candia, D., Cobos, Á., 2017. Whey protein film properties as affected by ultraviolet treatment under alkaline conditions. *International Dairy Journal* 73, 84–91. <https://doi.org/10.1016/j.idairyj.2017.05.009>.
- Dorgan, J.R., Lehermeier, H.J., Palade, L.-L., Cicero, J., 2001. Polylactides: properties and prospects of an environmentally benign plastic from renewable resources. *Macromolecular Symposia* 175, 55–66. [https://doi.org/10.1002/1521-3900\(200110\)175:1<55::AID-MASY55>3.0.CO;2-K](https://doi.org/10.1002/1521-3900(200110)175:1<55::AID-MASY55>3.0.CO;2-K).
- Durán, N., Lemes, A.P., Durán, M., Freer, J., Baeza, J., 2011. A minireview of cellulose nanocrystals and its potential integration as Co-product in bioethanol production. *Journal of the Chilean Chemical Society* 56, 672–677. <https://doi.org/10.4067/S0717-97072011000200011>.
- Elizabeth, R., 2006. Corn plastic to the rescue. *Smithsonian Magazine* 84–88.
- Emadian, S.M., Onay, T.T., Demirel, B., 2017. Biodegradation of bioplastics in natural environments. *Waste Management* 59, 526–536. <https://doi.org/10.1016/j.wasman.2016.10.006>.
- Fabra, M.J., Sánchez-González, L., Chiralt, A., 2014. Lysozyme release from isolate pea protein and starch based films and their antimicrobial properties. *Lebensmittel-Wissenschaft und -Technologie- Food Science and Technology* 55, 22–26. <https://doi.org/10.1016/j.lwt.2013.08.001>.
- Farah, S., Anderson, D.G., Langer, R., 2016. Physical and mechanical properties of PLA, and their functions in widespread applications — a comprehensive review. *Advanced Drug Delivery Reviews* 107, 367–392. <https://doi.org/10.1016/j.addr.2016.06.012>.
- Fennema, O., 1985. Chemical changes in food during processing—an overview. In: *Chemical Changes in Food During Processing*. Springer Netherlands, Dordrecht, pp. 1–16. [https://doi.org/10.1007/978-94-017-1016-9\\_1](https://doi.org/10.1007/978-94-017-1016-9_1).
- Finkielstein, S., Vournakis, J., September 22, 2015. Hemostatic compositions and therapeutic regimens. US 9139664 B2.
- Francis, R., Prestwich, G., Hunt, G., June 4, 2015. System and method of delivering a hyaluronic acid composition and a copper composition for treatment of dermatologic conditions. WO 2015081304 A1.
- Fu, Z., Wang, L., Li, D., Wei, Q., Adhikari, B., 2011. Effects of high-pressure homogenization on the properties of starch-plasticizer dispersions and their films. *Carbohydrate Polymers* 86, 202–207. <https://doi.org/10.1016/j.carbpol.2011.04.032>.
- Garrido, T., Etxabide, A., Guerrero, P., de la Caba, K., 2016. Characterization of agar/soy protein biocomposite films: effect of agar on the extruded pellets and compression moulded films. *Carbohydrate Polymers* 151, 408–416. <https://doi.org/10.1016/j.carbpol.2016.05.089>.
- Garside, M., 2019. Global Plastic Production 1950–2018. Statista. <https://www.statista.com/statistics/282732/global-production-of-plastics-since-1950/>.
- Generalic, E., 2019. “Starch” Croatian-English Chemistry Dictionary & Glossary. KTF-Split. <https://glossary.periodni.com/glossary.php?en&equals;starch>.
- Gennadios, A., 2004. Edible films and coatings from proteins. In: *Proteins in Food Processing*. <https://doi.org/10.1533/9781855738379.3.442>.
- Gewin, V., 2003. Genetically modified corn— environmental benefits and risks. *PLoS Biology* 1, e8. <https://doi.org/10.1371/journal.pbio.0000008>.
- Gigli, M., Fabbri, M., Lotti, N., Gamberini, R., Rimini, B., Munari, A., 2016. Poly(butylene succinate)-based polyesters for biomedical applications: a review. *European Polymer Journal* 75, 431–460. <https://doi.org/10.1016/j.eurpolymj.2016.01.016>.
- Gómez-Estaca, J., Montero, P., Gómez-Guillén, M.C., 2014. Shrimp (*Litopenaeus vannamei*) muscle proteins as source to develop edible films. *Food Hydrocolloids* 41, 86–94. <https://doi.org/10.1016/j.foodhyd.2014.03.032>.
- Gómez-Guillén, M.C., Giménez, B., López-Caballero, M.E., Montero, M.P., 2011. Functional and bioactive properties of collagen and gelatin from alternative sources: a review. *Food Hydrocolloids* 25, 1813–1827. <https://doi.org/10.1016/j.foodhyd.2011.02.007>.
- Gupta, P., Nayak, K.K., 2015. Characteristics of protein-based biopolymer and its application. *Polymer Engineering & Science* 55, 485–498. <https://doi.org/10.1002/pen.23928>.
- Gutiérrez, T.J., Tapia, M.S., Pérez, E., Famá, L., 2015. Structural and mechanical properties of edible films made from native and modified cush-cush yam and cassava starch. *Food Hydrocolloids* 45, 211–217. <https://doi.org/10.1016/j.foodhyd.2014.11.017>.
- Haghpanah, J.S., Yuvienco, C., Civay, D.E., Barra, H., Baker, P.J., Khapli, S., Voloshchuk, N., Gunasekar, S.K., Muthukumar, M., Montclare, J.K., 2009. Artificial protein block copolymers blocks comprising two distinct self-assembling domains. *ChemBioChem* 10, 2733–2735. <https://doi.org/10.1002/cbic.200900539>.
- Halimatul, M.J., Sapuan, S.M., Jawaid, M., Ishak, M.R., Ilyas, R.A., 2019a. Effect of sago starch and plasticizer

- content on the properties of thermoplastic films: mechanical testing and cyclic soaking-drying. *Polimery* 64, 32–41. <https://doi.org/10.14314/polimery.2019.6.5>.
- Halimatul, M.J., Sapuan, S.M., Jawaid, M., Ishak, M.R., Ilyas, R.A., 2019b. Water absorption and water solubility properties of sago starch biopolymer composite films filled with sugar palm particles. *Polimery* 64, 27–35. <https://doi.org/10.14314/polimery.2019.9.4>.
- Hassan, M.A., Yee, L.N., Yee, P.L., Ariffin, H., Raha, A.R., Shirai, Y., Sudesh, K., 2013. Sustainable production of polyhydroxyalkanoates from renewable oil-palm biomass. *Biomass and Bioenergy* 50, 1–9. <https://doi.org/10.1016/j.biombioe.2012.10.014>.
- Hazrol, M.D., Sapuan, S.M., Ilyas, R.A., Othman, M.L., Sherwani, S.F.K., 2020. Electrical properties of sugar palm nanocrystalline cellulose, reinforced sugar palm starch nanocomposites. *Polimery* 55, 33–40. <https://doi.org/10.14314/polimery.2020.5.4>.
- Hemsri, S., Asandei, A.D., Grieco, K., Parnas, R.S., 2011. Biopolymer composites of wheat gluten with silica and alumina. *Composites Part A: Applied Science and Manufacturing* 42, 1764–1773. <https://doi.org/10.1016/j.compositesa.2011.07.032>.
- Hietala, M., Mathew, A.P., Oksman, K., 2013. Bio-nanocomposites of thermoplastic starch and cellulose nanofibers manufactured using twin-screw extrusion. *European Polymer Journal* 49, 950–956. <https://doi.org/10.1016/j.eurpolymj.2012.10.016>.
- Hosseini, S.F., Rezaei, M., Zandi, M., Farahmandghavi, F., 2015. Fabrication of bio-nanocomposite films based on fish gelatin reinforced with chitosan nanoparticles. *Food Hydrocolloids* 44, 172–182. <https://doi.org/10.1016/j.foodhyd.2014.09.004>.
- Hu, X., Wang, X., Rnjak, J., Weiss, A.S., Kaplan, D.L., 2010. Biomaterials derived from silk–tropoelastin protein systems. *Biomaterials* 31, 8121–8131. <https://doi.org/10.1016/j.biomaterials.2010.07.044>.
- Ikada, Y., Tsuji, H., 2000. Biodegradable polyesters for medical and ecological applications. *Macromolecular Rapid Communications* 21, 117–132. [https://doi.org/10.1002/\(sici\)1521-3927\(20000201\)21:3<117::aid-marc117>3.3.co;2-o](https://doi.org/10.1002/(sici)1521-3927(20000201)21:3<117::aid-marc117>3.3.co;2-o).
- Ilyas, R.A., Sapuan, S.M., Ishak, M.R., Zainudin, E.S., 2017. Effect of delignification on the physical, thermal, chemical, and structural properties of sugar palm fibre. *BioResources* 12, 8734–8754. <https://doi.org/10.15376/biores.12.4.8734-8754>.
- Ilyas, R.A., Sapuan, S.M., 2020. The preparation methods and processing of natural fibre bio-polymer composites. *Current Organic Synthesis* 16, 1068–1070. <https://doi.org/10.2174/157017941608200120105616>.
- Ilyas, R.A., Sapuan, S.M., Atiqah, A., Ibrahim, R., Abrial, H., Ishak, M.R., Zainudin, E.S., Nurazzi, N.M., Atikah, M.S.N., Ansari, M.N.M., Asyraf, M.R.M., Supian, A.B.M., Ya, H., 2020a. Sugar palm (*Arenga pinnata* [Wurmb.] Merr) starch films containing sugar palm nanofibrillated cellulose as reinforcement: water barrier properties. *Polymer Composites* 41, 459–467. <https://doi.org/10.1002/pc.25379>.
- Ilyas, R.A., Sapuan, S.M., Ibrahim, R., Abrial, H., Ishak, M.R., Zainudin, E.S., Asrofi, M., Atikah, M.S.N., Huzaifah, M.R.M., Radzi, A.M., Azammi, A.M.N., Shaharuzaman, M.A., Nurazzi, N.M., Syafri, E., Sari, N.H., Norraahim, M.N.F., Jumaidin, R., 2019d. Sugar palm (*Arenga pinnata* (Wurmb.) Merr) cellulosic fibre hierarchy: a comprehensive approach from macro to nano scale. *Journal of Materials Research and Technology* 8, 2753–2766. <https://doi.org/10.1016/j.jmrt.2019.04.011>.
- Ilyas, R.A., Sapuan, S.M., Ibrahim, R., Abrial, H., Ishak, M.R., Zainudin, E.S., Atikah, M.S.N., Mohd Nurazzi, N., Atiqah, A., Ansari, M.N.M., Syafri, E., Asrofi, M., Sari, N.H., Jumaidin, R., 2019a. Effect of sugar palm nanofibrillated cellulose concentrations on morphological, mechanical and physical properties of biodegradable films based on agro-waste sugar palm (*Arenga pinnata* (Wurmb.) Merr) starch. *Journal of Materials Research and Technology* 8, 4819–4830. <https://doi.org/10.1016/j.jmrt.2019.08.028>.
- Ilyas, R.A., Sapuan, S.M., Ibrahim, R., Abrial, H., Ishak, M.R., Zainudin, E.S., Atiqah, A., Atikah, N., Syafri, E., Asrofi, M., Jumaidin, R., 2020b. Thermal, biodegradability and water barrier properties of bio-nanocomposites based on plasticised sugar palm starch and nanofibrillated celluloses from sugar palm fibres. *Journal of Biobased Materials and Bioenergy* 14, 1–13. <https://doi.org/10.1166/jbmb.2020.1951>.
- Ilyas, R.A., Sapuan, S.M., Ibrahim, R., Atikah, M.S.N., Atiqah, A., Ansari, M.N.M., Norraahim, M.N.F., 2019b. Production, processes and modification of nanocrystalline cellulose from agro-waste: a review. In: *Nanocrystalline Materials*. IntechOpen, pp. 3–32. <https://doi.org/10.5772/intechopen.87001>.
- Ilyas, R.A., Sapuan, S.M., Ishak, M.R., Zainudin, E.S., 2019c. Sugar palm nanofibrillated cellulose (*Arenga pinnata* (Wurmb.) Merr): effect of cycles on their yield, physico-chemical, morphological and thermal behavior. *International Journal of Biological Macromolecules* 123, 379–388. <https://doi.org/10.1016/j.ijbiomac.2018.11.124>.
- Ilyas, R.A., Sapuan, S.M., Ishak, M.R., 2018. Isolation and characterization of nanocrystalline cellulose from sugar palm fibres (*Arenga Pinnata*). *Carbohydrate Polymers* 181, 1038–1051. <https://doi.org/10.1016/j.carbpol.2017.11.045>.
- Ilyas, R.A., Sapuan, S.M., Ishak, M.R., Zainudin, E.S., 2018a. Development and characterization of sugar palm nanocrystalline cellulose reinforced sugar palm starch bionanocomposites. *Carbohydrate Polymers* 202, 186–202. <https://doi.org/10.1016/j.carbpol.2018.09.002>.
- Ilyas, R.A., Sapuan, S.M., Ishak, M.R., Zainudin, E.S., 2018b. Sugar palm nanocrystalline cellulose reinforced sugar palm starch composite: degradation and water-barrier properties. In: *IOP Conference Series: Materials Science and Engineering*. <https://doi.org/10.1088/1757-899X/368/1/012006>.
- Ilyas, R.A., Sapuan, S.M., Ishak, M.R., Zainudin, E.S., 2018e. Water transport properties of bio-nanocomposites reinforced by sugar palm (*arenga pinnata*) nanofibrillated

- cellulose. *Journal of Advanced Research in Fluid Mechanics and Thermal Sciences Journal* 51, 234–246.
- Ilyas, R.A., Sapuan, S.M., Ishak, M.R., Zainudin, E.S., Atikah, M.S.N., 2018c. Characterization of sugar palm nanocellulose and its potential for reinforcement with a starch-based composite. In: *Sugar Palm Biofibers, Biopolymers, and Biocomposites*, first ed. CRC Press/Taylor & Francis Group, Boca Raton, FL, pp. 189–220. <https://doi.org/10.1201/9780429443923-10>.
- Ilyas, R.A., Sapuan, S.M., Sanyang, M.L., Ishak, M.R., Zainudin, E.S., 2018d. Nanocrystalline cellulose as reinforcement for polymeric matrix nanocomposites and its potential applications: a review. *Current Analytical Chemistry* 14, 203–225. <https://doi.org/10.2174/1573411013666171003155624>.
- Jamaluddin, N., Razaina, M.T., Ishak, Z.M., 2016. Mechanical and morphology behaviours of polybutylene (succinate)/thermoplastic polyurethaneblend. *Procedia Chemistry* 19, 426–432. <https://doi.org/10.1016/j.proche.2016.03.034>.
- Jamshidian, M., Tehrani, E.A., Imran, M., Jacquot, M., Desobry, S., 2010. Poly-Lactic Acid: Production, Applications, Nanocomposites, and Release Studies. *Comprehensive Reviews in Food Science and Food Safety* 9, 552–571. <https://doi.org/10.1111/j.1541-4337.2010.00126.x>.
- Jiang, C., Wang, X., Gunawidjaja, R., Lin, Y.-H., Gupta, M.K., Kaplan, D.L., Naik, R.R., Tsukruk, V.V., 2007. Mechanical properties of robust ultrathin silk fibroin films. *Advanced Functional Materials* 17, 2229–2237. <https://doi.org/10.1002/adfm.200601136>.
- Johnson, E., Nichols, E., October 2000. High tap density chitosan, and methods of production. US 6130321 A.
- Jooyandeh, H., 2011. Whey protein films and coatings: a review. *Pakistan Journal of Nutrition* 10, 296–301. <https://doi.org/10.3923/pjn.2011.296.301>.
- Jumaidin, R., Ilyas, R.A., Saiful, M., Hussin, F., Mastura, M.T., 2019a. Water transport and physical properties of sugarcane bagasse fibre reinforced thermoplastic potato starch biocomposite. *Journal of Advanced Research in Fluid Mechanics and Thermal Sciences* 61, 273–281.
- Jumaidin, R., Khiruddin, M.A.A., Asyul Sutan Saidi, Z., Salit, M.S., Ilyas, R.A., 2020. Effect of cogon grass fibre on the thermal, mechanical and biodegradation properties of thermoplastic cassava starch biocomposite. *International Journal of Biological Macromolecules* 146, 746–755. <https://doi.org/10.1016/j.ijbiomac.2019.11.011>.
- Jumaidin, R., Saidi, Z.A.S., Ilyas, R.A., Ahmad, M.N., Wahid, M.K., Yaakob, M.Y., Maidin, N.A., Rahman, M.H.A., Osman, M.H., 2019b. Characteristics of cogon grass fibre reinforced thermoplastic cassava starch biocomposite: water absorption and physical properties. *Journal of Advanced Research in Fluid Mechanics and Thermal Sciences* 62 (62), 43–52.
- Karimi, S., Tahir, P., Dufresne, A., Karimi, A., Abdulkhani, A., 2014. A comparative study on characteristics of nanocellulose reinforced thermoplastic starch biofilms prepared with different techniques. *Nordic Pulp and Paper Research Journal* 29, 41–45.
- Kaushik, A., Singh, M., Verma, G., 2010. Green nanocomposites based on thermoplastic starch and steam exploded cellulose nanofibrils from wheat straw. *Carbohydrate Polymers* 82, 337–345. <https://doi.org/10.1016/j.carbpol.2010.04.063>.
- Khwaldia, K., Arab-Tehrany, E., Desobry, S., 2010. Biopolymer coatings on paper packaging materials. *Comprehensive Reviews in Food Science and Food Safety* 9, 82–91. <https://doi.org/10.1111/j.1541-4337.2009.00095.x>.
- Kinsella, J.E., Morr, C.V., 1984. Milk proteins: physicochemical and functional properties. *CRC Critical Reviews in Food Science & Nutrition* 21, 197–262. <https://doi.org/10.1080/10408398409527401>.
- Koutsopoulos, S., Unsworth, L.D., Nagai, Y., Zhang, S., 2009. Controlled release of functional proteins through designer self-assembling peptide nanofiber hydrogel scaffold. *Proceedings of the National Academy of Sciences* 106, 4623–4628. <https://doi.org/10.1073/pnas.0807506106>.
- Kvien, I., Sugiyama, J., Votrubeč, M., Oksman, K., 2007. Characterization of starch based nanocomposites. *Journal of Materials Science* 42, 8163–8171. <https://doi.org/10.1007/s10853-007-1699-2>.
- Lee, J.-H., Lee, J., Song, K.B., 2015a. Development of a chicken feet protein film containing essential oils. *Food Hydrocolloids* 46, 208–215. <https://doi.org/10.1016/j.foodhyd.2014.12.020>.
- Lee, J.-H., Won, M., Song, K.B., 2015b. Physical properties and antimicrobial activities of porcine meat and bone meal protein films containing coriander oil. *Lebensmittel-Wissenschaft und -Technologie- Food Science and Technology* 63, 700–705. <https://doi.org/10.1016/j.lwt.2015.03.043>.
- Li, X., Ji, N., Qiu, C., Xia, M., Xiong, L., Sun, Q., 2015. The effect of peanut protein nanoparticles on characteristics of protein- and starch-based nanocomposite films: a comparative study. *Industrial Crops and Products* 77, 565–574. <https://doi.org/10.1016/j.indcrop.2015.09.026>.
- Lim, S., Jane, J., 1994. Storage stability of injection-molded starch-zein plastics under dry and humid conditions. *Journal of Environmental Polymer Degradation* 2, 111–120. <https://doi.org/10.1007/BF02074779>.
- Liu, D., Zhong, T., Chang, P.R., Li, K., Wu, Q., 2010. Starch composites reinforced by bamboo cellulosic crystals. *Bioresource Technology* 101, 2529–2536. <https://doi.org/10.1016/j.biortech.2009.11.058>.
- Liu, L., Yang, J.-P., Ju, X.-J., Xie, R., Liu, Y.-M., Wang, W., Zhang, J.-J., Niu, C.H., Chu, L.-Y., 2011. Monodisperse core-shell chitosan microcapsules for pH-responsive burst release of hydrophobic drugs. *Soft Matter* 7, 4821. <https://doi.org/10.1039/c0sm01393e>.
- Llanos, J.H.R., Tadini, C.C., 2018. Preparation and characterization of bio-nanocomposite films based on cassava starch or chitosan, reinforced with montmorillonite or bamboo nanofibers. *International Journal of Biological Macromolecules* 107, 371–382. <https://doi.org/10.1016/j.ijbiomac.2017.09.001>.
- Lu, J., Tappel, R.C., Nomura, C.T., 2009. Mini-Review: biosynthesis of poly(hydroxyalkanoates). *Polymer Reviews* 49, 226–248. <https://doi.org/10.1080/15583720903048243>.

- Lu, Y., Weng, L., Cao, X., 2006. Morphological, thermal and mechanical properties of ramie crystallites—reinforced plasticized starch biocomposites. *Carbohydrate Polymers* 63, 198–204. <https://doi.org/10.1016/j.carbpol.2005.08.027>.
- Lu, Y., Weng, L., Cao, X., 2005. Biocomposites of plasticized starch reinforced with cellulose crystallites from cottonseed linter. *Macromolecular Bioscience* 5, 1101–1107. <https://doi.org/10.1002/mabi.200500094>.
- Lucchesi, L., Xie, H., December 8, 2015. Wound dressing devices and methods. US 9205170 B2.
- Ma, X., Qiao, C., Wang, X., Yao, J., Xu, J., 2019. Structural characterization and properties of polyols plasticized chitosan films. *International Journal of Biological Macromolecules* 135, 240–245. <https://doi.org/10.1016/j.ijbiomac.2019.05.158>.
- Martinez, K., Baeza, R., Millan, F., Pilosof, A., 2005. Effect of limited hydrolysis of sunflower protein on the interactions with polysaccharides in foams. *Food Hydrocolloids* 19, 361–369. <https://doi.org/10.1016/j.foodhyd.2004.10.002>.
- Mathew, A.P., Thielemans, W., Dufresne, A., 2008. Mechanical properties of nanocomposites from sorbitol plasticized starch and tunicin whiskers. *Journal of Applied Polymer Science* 109, 4065–4074. <https://doi.org/10.1002/app.28623>.
- McCarthy, S., Gregory, K., Wiesmann, W., Campbell, T., March 11, 2014. Wound dressing and method for controlling severe, life threatening bleeding. US 8668924 B2.
- McCarthy, S., McGrath, B., Winata, E., September 18, 2012. Absorbable tissue dressing assemblies, systems, and methods formed from hydrophilic polymer sponge structures such as chitosan. US 8269058 B2.
- Mchugh, T.H., Aujard, J.-F., Krochta, J.M., 1994. Plasticized whey protein edible films: water vapor permeability properties. *Journal of Food Science* 59, 416–419. <https://doi.org/10.1111/j.1365-2621.1994.tb06980.x>.
- McHugh, T.H., Krochta, J.M., 1994. Sorbitol- vs glycerol-plasticized whey protein edible films: integrated oxygen permeability and tensile property evaluation. *Journal of Agricultural and Food Chemistry* 42, 841–845. <https://doi.org/10.1021/jf00040a001>.
- Mehyar, G.F., Han, J.H., 2006. Physical and mechanical properties of high-amylose rice and pea starch films as affected by relative humidity and plasticizer. *Journal of Food Science* 69, E449–E454. <https://doi.org/10.1111/j.1365-2621.2004.tb09929.x>.
- Mhd Haniffa, M., Ching, Y., Abdullah, L., Poh, S., Chuah, C., 2016. Review of bionanocomposite coating films and their applications. *Polymers* 8, 246. <https://doi.org/10.3390/polym8070246>.
- Montero, B., Rico, M., Rodríguez-Llamazares, S., Barral, L., Bouza, R., 2017. Effect of nanocellulose as a filler on biodegradable thermoplastic starch films from tuber, cereal and legume. *Carbohydrate Polymers* 157, 1094–1104. <https://doi.org/10.1016/j.carbpol.2016.10.073>.
- Morin-Crini, N., Lichtfouse, E., Torri, G., Crini, G., 2019. Fundamentals and applications of chitosan. In: Crini, G., Lichtfouse, E. (Eds.), *Sustainable Agriculture Reviews, Sustainable Agriculture Reviews*. Springer International Publishing, Cham, pp. 49–123. [https://doi.org/10.1007/978-3-030-16538-3\\_2](https://doi.org/10.1007/978-3-030-16538-3_2).
- Müller, C.M.O., Laurindo, J.B., Yamashita, F., 2009. Effect of cellulose fibers addition on the mechanical properties and water vapor barrier of starch-based films. *Food Hydrocolloids* 23, 1328–1333. <https://doi.org/10.1016/j.foodhyd.2008.09.002>.
- Nasri-Nasrabadi, B., Behzad, T., Bagheri, R., 2014. Preparation and characterization of cellulose nanofiber reinforced thermoplastic starch composites. *Fibers and Polymers* 15, 347–354. <https://doi.org/10.1007/s12221-014-0347-0>.
- Nasseri, R., Mohammadi, N., 2014. Starch-based nanocomposites: a comparative performance study of cellulose whiskers and starch nanoparticles. *Carbohydrate Polymers* 106, 432–439. <https://doi.org/10.1016/j.carbpol.2014.01.029>.
- Nataraj, D., Sakkar, S., Meghwal, M., Reddy, N., 2018. Cross-linked chitosan films with controllable properties for commercial applications. *International Journal of Biological Macromolecules* 120, 1256–1264. <https://doi.org/10.1016/j.ijbiomac.2018.08.187>.
- Nazrin, A., Sapuan, S.M., Zuhri, M.Y.M., Ilyas, R.A., Syafiq, R., Sherwani, S.F.K., 2020. Nanocellulose Reinforced Thermoplastic Starch (TPS), Polylactic Acid (PLA), and Polybutylene Succinate (PBS) for Food Packaging Applications. *Frontiers in Chemistry* 8, 1–12. <https://doi.org/10.3389/fchem.2020.00213>.
- Norizan, M.N., Abdan, K., Ilyas, R.A., Biofibers, S.P., 2020. Effect of fiber orientation and fiber loading on the mechanical and thermal properties of sugar palm yarn fiber reinforced unsaturated polyester resin composites. *Polimery* 65, 34–43. <https://doi.org/10.14314/polimery.2020.2.5>.
- Noshirvani, N., Ghanbarzadeh, B., Fasihi, H., Almasi, H., 2016. Starch-PVA nanocomposite film incorporated with cellulose nanocrystals and MMT: a comparative study. *International Journal of Food Engineering* 12, 37–48. <https://doi.org/10.1515/ijfe-2015-0145>.
- Nurazzi, N.M., Khalina, A., Sapuan, S.M., Ilyas, R.A., Rafiqah, S.A., Hanafee, Z.M., 2019a. Thermal properties of treated sugar palm yarn/glass fiber reinforced unsaturated polyester hybrid composites. *Journal of Materials Research and Technology* 9 (2), 1606–1618. <https://doi.org/10.1016/j.jmrt.2019.11.086>.
- Nurazzi, N.M., Khalina, A., Sapuan, S.M., Ilyas, R.A., 2019b. Mechanical properties of sugar palm yarn / woven glass fiber reinforced unsaturated polyester composites : effect of fiber loadings and alkaline treatment. *Polimery* 64, 12–22. <https://doi.org/10.14314/polimery.2019.10.3>.
- Oryan, A., Sahvieh, S., 2017. Effectiveness of chitosan scaffold in skin, bone and cartilage healing. *International Journal of Biological Macromolecules* 104, 1003–1011. <https://doi.org/10.1016/j.ijbiomac.2017.06.124>.
- Oussalah, M., Caillet, S., Salmiéri, S., Saucier, L., Lacroix, M., 2004. Antimicrobial and antioxidant effects of milk protein-based film containing essential oils for the preservation of whole beef muscle. *Journal of Agricultural and Food Chemistry* 52, 5598–5605. <https://doi.org/10.1021/jf049389q>.
- Pang, X., Zhuang, X., Tang, Z., Chen, X., 2010. Polylactic acid (PLA): research, development and industrialization. *Biotechnology Journal* 5, 1125–1136. <https://doi.org/10.1002/biot.201000135>.

- Park, H.J., Kim, S.H., Lim, S.T., Shin, D.H., Choi, S.Y., Hwang, K.T., 2000. Grease resistance and mechanical properties of isolated soy protein-coated paper. *Journal of the American Oil Chemists' Society* 77, 269–273. <https://doi.org/10.1007/s11746-000-0044-2>.
- Patricia, P., Norma, A.S., Guzm, D.L., Fripiat, J.J., 2007. In: *Mesoporous Silica From Rice Hull Ash*, vol. 619, pp. 614–619. <https://doi.org/10.1002/jctb>.
- Pérez-Arauz, A.O., Aguilar-Rabiela, A.E., Vargas-Torres, A., Rodríguez-Hernández, A.-I., Chavarría-Hernández, N., Vergara-Porras, B., López-Cuellar, M.R., 2019. Production and characterization of biodegradable films of a novel polyhydroxyalkanoate (PHA) synthesized from peanut oil. *Food Packaging and Shelf Life* 20, 100297. <https://doi.org/10.1016/j.foodpack.2019.01.001>.
- Pérez-Gago, M.B., Rhim, J.-W., 2014. Edible coating and film materials. In: *Innovations in Food Packaging*. Elsevier, pp. 325–350. <https://doi.org/10.1016/B978-0-12-394601-0.00013-8>.
- Pighinelli, L., 2019. Methods of chitin production a short review. *American Journal of Biomedical Science & Research* 3, 307–314. <https://doi.org/10.34297/AJBSR.2019.03.000682>.
- Pol, H., Dawson, P., Acton, J., Ogale, A., 2002. Soy protein isolate/corn-zein laminated films: transport and mechanical properties. *Journal of Food Science* 67, 212–217. <https://doi.org/10.1111/j.1365-2621.2002.tb11386.x>.
- Pothakamury, U.R., Barbosa-Cánovas, G.V., 1995. Fundamental aspects of controlled release in foods. *Trends in Food Science & Technology* 6, 397–406. [https://doi.org/10.1016/S0924-2244\(00\)89218-3](https://doi.org/10.1016/S0924-2244(00)89218-3).
- Qiu, Y., Park, K., 2001. Environment-sensitive hydrogels for drug delivery. *Advanced Drug Delivery Reviews* 53, 321–339. [https://doi.org/10.1016/S0169-409X\(01\)00203-4](https://doi.org/10.1016/S0169-409X(01)00203-4).
- Rangavajihyala, N., Ghorpade, V., Hanna, M., 1997. Solubility and molecular properties of heat-cured soy protein films. *Journal of Agricultural and Food Chemistry* 45, 4204–4208. <https://doi.org/10.1021/jf9702048>.
- Rasal, R.M., Janorkar, A.V., Hirt, D.E., 2010. Poly(lactic acid) modifications. *Progress in Polymer Science* 35, 338–356. <https://doi.org/10.1016/j.progpolymsci.2009.12.003>.
- Rhim, J.-W., Lee, J.-H., Hong, S.-I., 2006. Water resistance and mechanical properties of biopolymer (alginate and soy protein) coated paperboards. *Lebensmittel-Wissenschaft und Technologie- Food Science and Technology* 39, 806–813. <https://doi.org/10.1016/j.lwt.2005.05.008>.
- Rhim, J.W., Weller, C.L., Gennadios, A., 2004. Effects of soy protein coating on shell strength and quality of shell eggs. *Food Science and Biotechnology* 13 (4), 455–459.
- Rocha, M. da, Loiko, M.R., Tondo, E.C., Prentice, C., 2014. Physical, mechanical and antimicrobial properties of Argentine anchovy (*Engraulis anchoita*) protein films incorporated with organic acids. *Food Hydrocolloids* 37, 213–220. <https://doi.org/10.1016/j.foodhyd.2013.10.017>.
- Rochman, C.M., Browne, M.A., Halpern, B.S., Hentschel, B.T., Hoh, E., Karapanagioti, H.K., Rios-Mendoza, L.M., Takada, H., Teh, S., Thompson, R.C., 2013. Classify plastic waste as hazardous. *Nature* 494, 169–171. <https://doi.org/10.1038/494169a>.
- Rokhade, A.P., Shelke, N.B., Patil, S.A., Aminabhavi, T.M., 2007. Novel interpenetrating polymer network microspheres of chitosan and methylcellulose for controlled release of theophylline. *Carbohydrate Polymers* 69, 678–687. <https://doi.org/10.1016/j.carbpol.2007.02.008>.
- Romani, V.P., Hernández, C.P., Martins, V.G., 2018. Pink pepper phenolic compounds incorporation in starch/protein blends and its potential to inhibit apple browning. *Food Packaging and Shelf Life* 15, 151–158. <https://doi.org/10.1016/j.foodpack.2018.01.003>.
- Roy, J.C., Salaün, F., Giraud, S., Ferri, A., Chen, G., Guan, J., 2017. Solubility of chitin: solvents, solution behaviors and their related mechanisms. In: *Solubility of Polysaccharides*. InTech. <https://doi.org/10.5772/intechopen.71385>.
- Sahay, N., Ierapetritou, M., 2009. Nihar SCM. IFAC Proceedings Volumes (IFAC-PapersOnline) 7, 405–410. <https://doi.org/10.1002/aic>.
- Sai-Ut, S., Benjakul, S., Rawdkuen, S., 2015. Retardation of lipid oxidation using gelatin film incorporated with longan seed extract compared with BHT. *Journal of Food Science & Technology* 52, 5842–5849. <https://doi.org/10.1007/s13197-014-1631-0>.
- Salgado, P.R., López-Caballero, M.E., Gómez-Guillén, M.C., Mauri, A.N., Montero, M.P., 2013. Sunflower protein films incorporated with clove essential oil have potential application for the preservation of fish patties. *Food Hydrocolloids* 33, 74–84. <https://doi.org/10.1016/j.foodhyd.2013.02.008>.
- Sanyang, M.L., Ilyas, R.A., Sapuan, S.M., Jumaidin, R., 2018. Sugar palm starch-based composites for packaging applications. In: *Bionanocomposites for Packaging Applications*. Springer International Publishing, Cham, pp. 125–147. [https://doi.org/10.1007/978-3-319-67319-6\\_7](https://doi.org/10.1007/978-3-319-67319-6_7).
- Sanyang, M.L., Sapuan, S.M., Jawaaid, M., Ishak, M.R., Sahari, J., 2016. Recent developments in sugar palm (*Arenga pinnata*) based biocomposites and their potential industrial applications: a review. *Renewable and Sustainable Energy Reviews* 54, 533–549. <https://doi.org/10.1016/j.rser.2015.10.037>.
- Sawyer, D.J., 2003. Bioprocessing—no longer a field of dreams. *Macromolecular Symposia* 201, 271–282. <https://doi.org/10.1002/masy.200351130>.
- Schmid, M., Müller, K., 2019. Whey protein-based packaging films and coatings. *Whey Proteins*. <https://doi.org/10.1016/b978-0-12-812124-5.00012-6>.
- Shah, A.A., Hasan, F., Hameed, A., Ahmed, S., 2008. Biological degradation of plastics: a comprehensive review. *Biotechnology Advances* 26, 246–265. <https://doi.org/10.1016/j.biotechadv.2007.12.005>.
- Sharma, L., Singh, C., 2016. Sesame protein based edible films: development and characterization. *Food Hydrocolloids* 61, 139–147. <https://doi.org/10.1016/j.foodhyd.2016.05.007>.
- Shi, W., Dumont, M.-J., 2014. Processing and physical properties of canola protein isolate-based films. *Industrial Crops and Products* 52, 269–277. <https://doi.org/10.1016/j.indcrop.2013.10.037>.
- Siakeng, R., Jawaaid, M., Ariffin, H., Sapuan, S.M., Asim, M., Saba, N., 2019. Natural fiber reinforced polylactic acid



- composites: a review. *Polymer Composites* 40, 446–463. <https://doi.org/10.1002/pc.24747>.
- Siew, D.C.W., Heilmann, C., Easteal, A.J., Cooney, R.P., 1999. Solution and film properties of sodium caseinate/glycerol and sodium caseinate/polyethylene glycol edible coating systems. *Journal of Agricultural and Food Chemistry* 47, 3432–3440. <https://doi.org/10.1021/jf9806311>.
- Singhvi, M.S., Zinjarde, S.S., Gokhale, D.V., 2019. Polylactic acid: synthesis and biomedical applications. *Journal of Applied Microbiology* 127, 1612–1626. <https://doi.org/10.1111/jam.14290>.
- Slavutsky, A.M., Bertuzzi, M.A., 2014. Water barrier properties of starch films reinforced with cellulose nanocrystals obtained from sugarcane bagasse. *Carbohydrate Polymers* 110, 53–61. <https://doi.org/10.1016/j.carbpol.2014.03.049>.
- Sobral, P.J.A., Menegalli, F.C., Hubinger, M.D., Roques, M.A., 2001. Mechanical, water vapor barrier and thermal properties of gelatin based edible films. *Food Hydrocolloids* 15, 423–432. [https://doi.org/10.1016/S0268-005X\(01\)00061-3](https://doi.org/10.1016/S0268-005X(01)00061-3).
- Song, H., Lee, S.Y., 2006. Production of succinic acid by bacterial fermentation. *Enzyme and Microbial Technology* 39, 352–361. <https://doi.org/10.1016/j.enzmictec.2005.11.043>.
- Spaccini, R., Todisco, D., Drosos, M., Nebbioso, A., Piccolo, A., 2016. Decomposition of bio-degradable plastic polymer in a real on-farm composting process. *Chemical and Biological Technologies in Agriculture* 3 (1), 4. <https://doi.org/10.1186/s40538-016-0053-9>.
- Stuchell, Y.M., Krochta, J.M., 1994. Enzymatic treatments and thermal effects on edible soy protein films. *Journal of Food Science* 59, 1332–1337. <https://doi.org/10.1111/j.1365-2621.1994.tb14709.x>.
- Su, J.-F., Huang, Z., Yuan, X.-Y., Wang, X.-Y., Li, M., 2010. Structure and properties of carboxymethyl cellulose/soy protein isolate blend edible films crosslinked by Maillard reactions. *Carbohydrate Polymers* 79, 145–153. <https://doi.org/10.1016/j.carbpol.2009.07.035>.
- Syafri, E., Kasim, A., Abral, H., Sudirman, Sulungbudi, G.T., Sanjay, M.R., Sari, N.H., 2018. Synthesis and characterization of cellulose nanofibers (CNF) ramie reinforced cassava starch hybrid composites. *International Journal of Biological Macromolecules* 120, 578–586. <https://doi.org/10.1016/j.ijbiomac.2018.08.134>.
- Syafri, E., Sudirman, M., Yulianti, E., Deswita, Asrofi, M., Abral, H., Sapuan, S.M., Ilyas, R.A., Fudholi, A., 2019. Effect of sonication time on the thermal stability, moisture absorption, and biodegradation of water hyacinth (*Eichhornia crassipes*) nanocellulose-filled bengkuang (*Pachyrhizus erosus*) starch biocomposites. *Journal of Materials Research and Technology* 8, 6223–6231. <https://doi.org/10.1016/j.jmrt.2019.10.016>.
- Tan, Z., Yi, Y., Wang, H., Zhou, W., Yang, Y., Wang, C., 2016. Physical and degradable properties of mulching films prepared from natural fibers and biodegradable polymers. *Applied Sciences* 6, 147. <https://doi.org/10.3390/app6050147>.
- Teixeira, E. de M., Pasquini, D., Curvelo, A.A.S.S., Corradini, E., Belgacem, M.N., Dufresne, A., 2009. Cassava bagasse cellulose nanofibrils reinforced thermoplastic cassava starch. *Carbohydrate Polymers* 78, 422–431. <https://doi.org/10.1016/j.carbpol.2009.04.034>.
- Teixeira, E.D.M., Lotti, C., Corrêa, A.C., Teodoro, K.B.R., Marconcini, J.M., Mattoso, L.H.C., 2011. Thermoplastic corn starch reinforced with cotton cellulose nanofibers. *Journal of Applied Polymer Science* 120, 2428–2433. <https://doi.org/10.1002/app.33447>.
- Thiagamani, S.M.K., Rajini, N., Siengchin, S., Varada Rajulu, A., Hariram, N., Ayrilmis, N., 2019. Influence of silver nanoparticles on the mechanical, thermal and antimicrobial properties of cellulose-based hybrid nanocomposites. *Composites Part B: Engineering* 165, 516–525. <https://doi.org/10.1016/j.compositesb.2019.02.006>.
- Thunwall, M., Boldizar, A., Rigdahl, M., 2006. Compression molding and tensile properties of thermoplastic potato starch materials. *Biomacromolecules* 7, 981–986. <https://doi.org/10.1021/bm050804c>.
- Tian, H., Xu, G., Yang, B., Guo, G., 2011. Microstructure and mechanical properties of soy protein/agar blend films: effect of composition and processing methods. *Journal of Food Engineering* 107, 21–26. <https://doi.org/10.1016/j.jfoodeng.2011.06.008>.
- Tonyali, B., Cikriki, S., Oztop, M.H., 2018. Physicochemical and microstructural characterization of gum tragacanth added whey protein based films. *Food Research International* 105, 1–9. <https://doi.org/10.1016/j.foodres.2017.10.071>.
- Torchilin, V.P., 2005. Recent advances with liposomes as pharmaceutical carriers. *Nature Reviews Drug Discovery* 4, 145–160. <https://doi.org/10.1038/nrd1632>.
- UNEP, 2018. Single-use Plastics: A Roadmap for Sustainability. United Nation Environment Programme.
- Vachon, C., Yu, H.-L., Yefsah, R., Alain, R., St-Gelais, D., Lacroix, M., 2000. Mechanical and structural properties of milk protein edible films cross-linked by heating and  $\gamma$ -irradiation. *Journal of Agricultural and Food Chemistry* 48, 3202–3209. <https://doi.org/10.1021/jf991055r>.
- Vidal, R.R.L., Moraes, J.S., 2019. Removal of organic pollutants from wastewater using chitosan: a literature review. *International Journal of Environmental Science and Technology* 16, 1741–1754. <https://doi.org/10.1007/s13762-018-2061-8>.
- Wang, H., Hu, D., Ma, Q., Wang, L., 2016. Physical and antioxidant properties of flexible soy protein isolate films by incorporating chestnut (*Castanea mollissima*) bur extracts. *Lebensmittel-Wissenschaft und -Technologie- Food Science and Technology* 71, 33–39. <https://doi.org/10.1016/j.lwt.2016.03.025>.
- Xu, J., Guo, B.H., 2010. Poly(butylene succinate) and its copolymers: research, development and industrialization. *Biotechnology Journal* 5, 1149–1163. <https://doi.org/10.1002/biot.201000136>.
- Yang, S., Tang, Y., Wang, J., Kong, F., Zhang, J., 2014. Surface treatment of cellulosic paper with starch-based composites reinforced with nanocrystalline cellulose. *Industrial & Engineering Chemistry Research* 53, 13980–13988. <https://doi.org/10.1021/ie502125s>.
- Yoo, S., Krochta, J.M., 2011. Whey protein-polysaccharide blended edible film formation and barrier, tensile, thermal and transparency properties. *Journal of the Science of Food*

- and Agriculture 91, 2628–2636. <https://doi.org/10.1002/jsfa.4502>.
- Zhang, C., Guo, K., Ma, Y., Ma, D., Li, X., Zhao, X., 2010. Original article: incorporations of blueberry extracts into soybean-protein-isolate film preserve qualities of packaged lard. *International Journal of Food Science and Technology* 45, 1801–1806. <https://doi.org/10.1111/j.1365-2621.2010.02331.x>.
- Zhang, Y.-Q., Tao, M.-L., Shen, W.-D., Zhou, Y.-Z., Ding, Y., Ma, Y., Zhou, W.-L., 2004. Immobilization of L-asparaginase on the microparticles of the natural silk sericin protein and its characters. *Biomaterials* 25, 3751–3759. <https://doi.org/10.1016/j.biomaterials.2003.10.019>.
- Zhang, Y., Han, J.H., 2006. Mechanical and thermal characteristics of pea starch films plasticized with monosaccharides and polyols. *Journal of Food Science* 71, E109–E118. <https://doi.org/10.1111/j.1365-2621.2006.tb08891.x>.
- Zhong, N., Yuan, Q., 2013. Preparation and properties of molded blends of wheat gluten and cationic water-borne polyurethanes. *Journal of Applied Polymer Science* 128, 460–469. <https://doi.org/10.1002/app.38198>.
- Zhu, K.J., Hendren, R.W., Jensen, K., Pitt, C.G., 1991. Synthesis, properties, and biodegradation of poly(1,3-trimethylene carbonate). *Macromolecules* 24, 1736–1740. <https://doi.org/10.1021/ma00008a008>.
- Zuo, M., Song, Y., Zheng, Q., 2009. Preparation and properties of wheat gluten/methylcellulose binary blend film casting from aqueous ammonia: a comparison with compression molded composites. *Journal of Food Engineering* 91, 415–422. <https://doi.org/10.1016/j.jfoodeng.2008.09.019>.